

US009476058B2

# (12) United States Patent Lim

#### (54) METHOD FOR SPEEDING UP PLANT GROWTH AND IMPROVING YIELD BY INTRODUCING PHOSPHATASES IN TRANSGENIC PLANT

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(73) Assignee: Versitech Limited, Hong Kong (HK)

(\*) Notice: Subject to any disclaimer, the term of this

patent is extended or adjusted under 35

U.S.C. 154(b) by 615 days.

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(22) Filed: **Apr. 15, 2013** 

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#### Related U.S. Application Data

- (62) Division of application No. 12/640,674, filed on Dec. 17, 2009, now abandoned.
- (60) Provisional application No. 61/138,918, filed on Dec. 18, 2008.
- (51) **Int. Cl.**  *C12N 15/82* (2006.01) *C12N 9/16* (2006.01)
- (52) **U.S. Cl.** CPC ...... *C12N 15/8245* (2013.01); *C12N 9/16*
- (58) Field of Classification Search

See application file for complete search history.

(2013.01); C12N 15/8261 (2013.01)

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#### (57) ABSTRACT

Transgenic plants having increased growth rate, increased sugar content, and increase yield are disclosed, and methods for making the same. The transgenic plants have a gene coding for a phosphatase having a C-terminal motif under control of a heterologous promoter incorporated into the genomic DNA of the plant.

#### 10 Claims, 14 Drawing Sheets

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U.S. Appl. No. 14/945,406, filed Nov. 18, 2015, Lim.

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FIG. 1

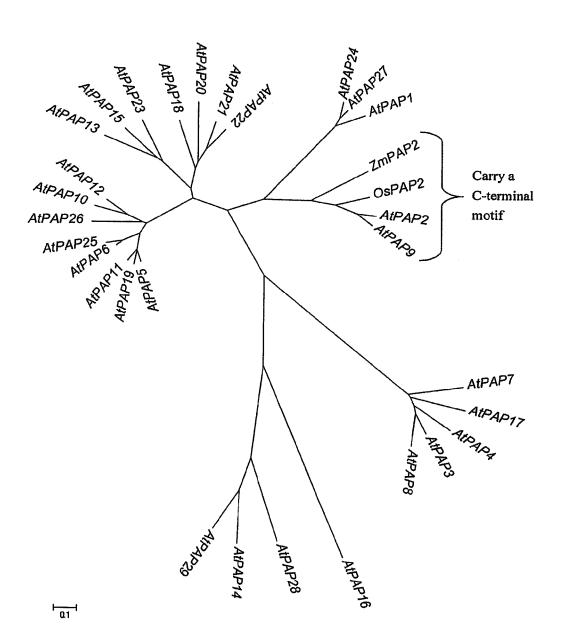


FIG. 2A

Protein Sequences (A) Transmamhrana	/ 'C / A !!   1\
The transmembrane	motif (Aligned)
AtPAP2_gi 15222978 ref NP_172843	TOTAL A LMVV VLL FII FF
AtPAP9_gi 20257481 gb AAM15910	LEXI BANVAVOVIE EXECUTE
Brassica_rapa_subsppekinensis_clone	LNEVERLLUVIEVLLEFIIEFF
Hordeum_vulgare_PUT-161a-Hordeum_vulg	INCE IN VMFALMLEFAL FL
Medicago_AC202582_HTG_Medicago_trunca	MAN AND THE PROPERTY OF THE PARTY OF THE PAR
OSPAP2_NM_001065273	LE LLIVE VMFALVL ELLEL
Poplar_trichocarpa_ref[NC_008476.1	AND A VLVL AFVER LEYA
Saccharum_officinarum_28138_PlantGDB	LYLISOVLFALLLEFFFFL
Solanum_tuberosum_PUT-157a-Solanum_tu	NO VENTER MIVEFL
Vitis_viniferaAM458569_modified	NO CAPACITAL AFMENUTER
Zea_mays_EU975503	LYLLIGOVMFALLLEFFEIL
Physcomitrella_patens_subsppatens_g	DIVAFLEVLALE CHAAALE
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AtPAP2_gi[15222978]ref[NP_172843 AtPAP9_gi[20257481]gb[AAM15910 Brassica rapa subsp. pekinensis_clone Hordeum_vulgare_PUT-161a-Hordeum_vulg Medicago_AC202582_HTG_Medicago_trunca OSPAP2_NM_001065273 Poplar_trichocarpa_ref[NC_008476.1]	EST IN SOME VENT OF STREET OF THE STREET OF
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### FIG. 2B

AtPAP2		LLLEWSVFVS	SAUSKATUSI	E NAL NRS GD	SKKIGMSOVD	SPSDLDWLÖE	58
BnPAP2 GmPAP2	MINDER -E - F	ILEEUSVFIIS	SANAKATLE		SEE I KWSHVD	SPSDLDWLGI	
ZmPAP2		LEGLENNFEH	PAESKEELIA	MP IIT L ENS GA 日記 - TAISES GN	TYNERWSONE DUK I HWSOER	SPSDLDELA I	
ALPAP 15	MICCOG-8-F	LLLLUEGFUS	PAUSSABEE	66 - 656666		EEBESEEEE	19
Consensus		LLXXXXXFXX	XXXXAXXXX	XPXXXXXSGX	XXXXXWSXXX	XPXXLDXXXX	
AtPAP2		FIGYKELNES	STWEEGEGEN	SLPLT-NLRS	NYTERIFRWS	ESEIDPKHKD	117
BnPAP2 GmPAP2		FIGYEFLNES	ETWOMOS GAN ATWRINGS GNU		NYTERIFRWT NYEFRIFEWT	SEINPKHKD	116
ZmPAP2		FUGYUFLNUS	ASWREGS GED		EYGERLERWE	がRERSWRHND MSEINSKEOD	118
ALPAP15		PYEGGGGGG	EERGEESEV			SOATOURDED	
Consensus		FXGYXFLXXS	XXWXXGXGXX		XYXFRXFXWX	XXEIXXXXXD	,
AIPAP2			FGS-GYGMPE	QIHLEFEN	- MV NIMR VMF	VAGDGE	
BnPAP2 GmPAP2		HLLAESEQVØ HLLAESEEVS	FOSAGNORPE FASHE OPG	QIHLAFED	-KVNKMRVIIF	VAGDGE	
ZmPAP2		HRYAYSADVE	VGDEA - RPE	QUHLAFAD	- EVDEMRVUS	WARDER VEGDRE	
ALPAP15		RRYEREBUIG	FEE-BEPE	OIELEUSEDH	- REGRIMARM	MICEE GROKK	
Consensus		HXXAXSXXXX	FXXXXVGXPE	QIHLXFXXXH	-XXXXMRVXX	XXGDXX	
AtPAP2 BnPAP2	E	RHVRYGERK-	-DULENBAAA		CDEPANSTILG	WRDPGWIFDT	218
GmPAP2		REVRYCEDK - 可以VRYCERE -	-DELANSAAA -DKLDGNAVA		CMAPANSTVC CDAPANESVG		
ZmPAP2	E	RYVRYGEGKE	DOKEWKEYGT		COWPANSEVA	WROPGE INDE	
	arendeledh	RANGEOUTH	-ESCEHENKO		CDWPANSEVE EDGEDNMTES	BEERCHIBHY	145
Consensus	· · · · · · · · · · · · · · · · · · ·	XXVRYGXXXX	-DXLXXXAXX	XXXXXEXXHW	CDXPANXXXX	WRDPGXIXXX	
AtPAP2	VMKNLNDGVR VMKNLNDGVR	YYYDVGSDSK	-CWSEIHSNI		A FMFGDM	GGAMPYMTFI	274
GmPAP2		AAAMAGNONG AAAMAGSOSK	- GWSEIHSFI	ERNED SEET!	A FMFGDM A FUFGDM	GEAMPYNTFI	
ZmPAP2	MMKGLEPGER	YEYKVOSDIO	-GWSEINSFI	SROSEASETN	A FOFGDM	GENERAL ANTER	276
	BETOLKPETE	YYYKGGDESK	REMSETHER		EGRUAYVOOU	GERREYNTES	
	XXXXLXXGXR	AAAXAGXDXX	-GWSXIXSFX	XXXXXXXXXXX	A FXFGDM	GXXXPYNTXX	
AtPAP2 BnPAP2	RTQDESISTV RTQDESISTV	KWILRDIEAL	GDKPAMISHI	GDISYAR		• • • • • • • • •	
GmPAP2	RTQDESISTM	KWILRDIEAL KWILRDWEAL	GDKPA提送5HI GDKPAEK5HI	GDISYAR			311
ZmPAP2	RTOSESEST	KWILRDIEAL	GDKPAFIBHI	GD I SYAR		••••••	
AlPAP 15	EBENEW: SHO	Beebeeren	HWELDHIEDI	CDE2 A FEEE E	INGLEEDCKE	CSECEDENHE	249
Consensus	MIGALOTOTA	KWILRDXEAL	GDKPAXXSHI	GDISYAR	• • • • • • • • • • •		
AIPAP2 BRPAP2	GYSWVWDEFF GYSWVWDEFF	AQUEPIASNY AQIEPIASNY	PYHVCIGNHE	YDESTOPWEP	DWAASHYGND	GGGECGVPYS	
GmPAP2		AQIEPYASQV	BYHVC 1 GNHE	YDEPTQPWKP YDWP <u>U</u> QPWKP	DWGT YGND	GGGECGVPYS GGGECGVPYS	369 372
ZmPAP2	GYSWVWYHFF	SCIEPIANN	PYHVCIGNHE	YDWPSQPWKP	WWAT - YOUD	GGGECGNPYS	370
AIPAP 15	Jan will trad dett	REMEMBILE RA	<b>Ь ∏Ņ∧∭Ē</b> ONHE	REPOSERER	BEES BEES	MENERAYS	293
A1PAP2	GYSWXWDXFF	XQXEPXASKV	BAHAC I GNHE	YDXXXQPWKP	XWX X • • YGX D	GGGECGXPYS	
	LKENMPGNSS LKENMPGNSS	EETG-MKAPP	TRNLYYS黨DM		ETNFTKGGEO	YEFWKRDLES	
	LRFNMPGNSS	ELTGNAAAPP	TRNLYYSFDM		ETNFUKCORO ETNFUKCORO	YEFEKROLES YEFEKHOLES	
ZmPAP2	SREEMP ON SIL	LETCH - GGPD	TRNLYYSFD	GYVHFVYMST	ETHFYGGEE	HAFLKADLEK	429
AlPAP 15 Consensus	XXFXMPGNSS		BEELYVSFNA	CCHHFVMLGA	KINKEREO	AEMIKKOLKK	
	VEREKTPEVV	VQGHRPMYTT	TRNLYYSXDX	GXVHFVYXST	ETNFXXCXXO	YXFXKXDLEX	
	VNREKTPFVV	VOGHRPMYTT	-SNEVRD版例页 -SNEVRDAMI	ROKMYEHLEP Rokmyehlep	LEVENNYTLA LEVENNYTLA	LWGHVHRYER	
GmPAP2	VNRSKTPFVV	VQGHRPMYTT	-SHENRDARE	REKMEEHLEP	LLVNNNVTLA	LWGHVHRYER LWGHVHRYER	
ZmPAP2 AtPAP15	VNRSRTPFVV	ECCHRPMYTE	-SDETTROAKU	ROOMEONLEP	LLV製器NVTLA	LWGHVHRYER	488
Consensus	V DR S DT PWDV	X Q G H R P MY T X	*SXEXRDAXX	REGWRESWEE	r r z z z g z d z d z z		402
AtPAP2	FCP I SNNT	-ccTcW		XXXMXEXLEP	LXVXXNVTLA	LWGHVHRYER	
8nPAP2	FCPISNNT	-ceRoM	· · · · · · · · · · · · · · · · · · ·	VIGMGGGDWQ	PEWQ	··PRPNHPDE ··PRPNHPEE	
GmPAP2	FCPENNET	-ccknkghna	<b>GDKKCKU</b> AHD	VIGMAGODWO	PWW		
ZmPAP2 AlPAP15	ENERTHATEID ECHNINZG	CANTE-REE	EDZECKA ANT	A I CWCCODMO	PWQ	- PRPOHPOV	535
	FCPXXNXX	-CGXXX-XXX		VIGMXGQDWQ	PXWX		449
	P   F	********	POPEOSMYRM	GEFGYTRLVA	NKEKLTW-SE	· · PRPXHPXX	E7 ·
	PIF	• • • • • • • • • • • • • • • • • • • •	POPEOSMYR	GEFGYTRLVA	KEKLTK-SE	VGNHDG EV	
GmPAP2 ZmPAP2	PIF	*********	POPKWSTYRO	GEFGYTRLVA	近代質化し窓し・5条	VONHDG EV	680
ALPAP15	PYMEGECKWN	ETEERKECWO	P Q P E R SMY R C R Q P D M S A U R E	GEFGYKRLVA ENFCHQIILEM	ENEXWALWIN	NENDDEEEEV ACNHDG・・GA	575 609
Consensus	P   F	**********	POPXXSXYRX	GEFGYXRLVA	XXEK LXX - XX		209
AIPAP2	HDMVEMLASG	XV:5GEKEST	KEENOKEVPR	SATLMGKS		MOLMVMGVLL	629
BnPAP2	HOSVEILASG	EVISCRKEET	· · · · · · · · · · · · · · · · · · ·	SATLIGKE	ESDYLWYVKG	BOLLVBOVLL	623
ZmPAP2	HDMYE! EGSC	FAMECAGGG	ERVES			西東京 L 京原 C 東京図	633
AIPAP15	SPONK XEGE	DESECHBER		ezreber Ukrolozeta	のひにあなる 1 音を楽	GCSKMEKULL GCBRANGERO	529
Consensus	HDXXEIXXSG	XXXXXXXXX	· · · · XXXXXX	XXXXXCXXXV	XXXXXXXXXX	XXXXXXXXX	
AIPAP2	GFMMGFFTRG	KK-BSEENR	WIPVKNEET	657			
DIMPAPE	CERROLLING	<b>ルドログラ製製を開放</b> し	W選PVKNEET流	653 563			
ZMPAP?	CERMCHECAR	KKENACE - DO	化化分分化 计自己分级	oee			
Atpap16	<u> </u>	ES-3226-93	EEEEEENHCA	533			
Consensus	GXXXGXXXXX	XKXXXXXXX	MXXAKXEEXX				

FIG. 3

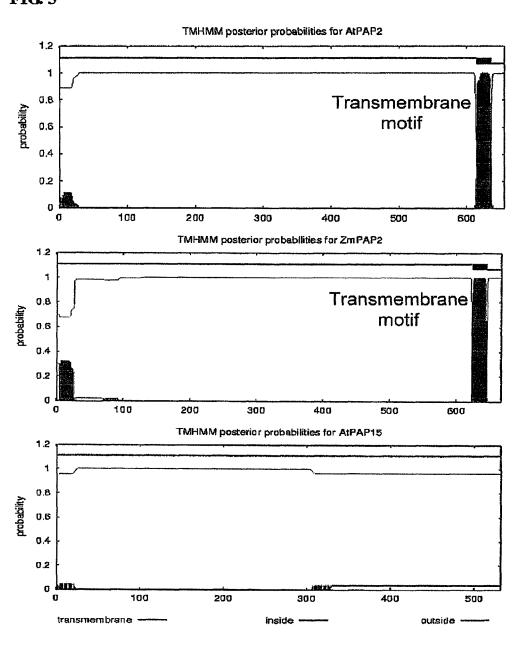
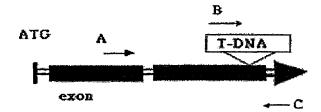


FIG. 4



atpap2-8 structure

## (a) Genomic PCR

WTatpap2-8

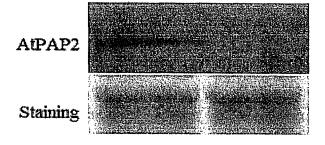


## (b) RT-PCR

WT atpap2-8 AtPAP2 EF

## (c) Western blotting

WT atpap2-8



**FIG.** 5

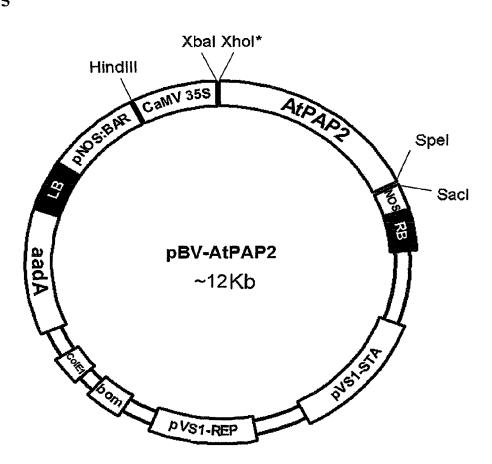
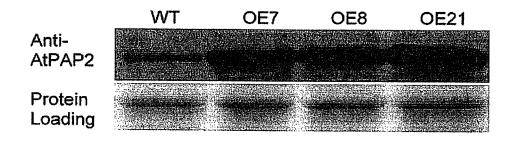
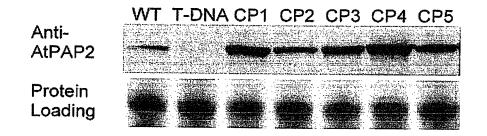
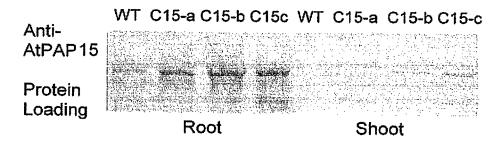


FIG. 6

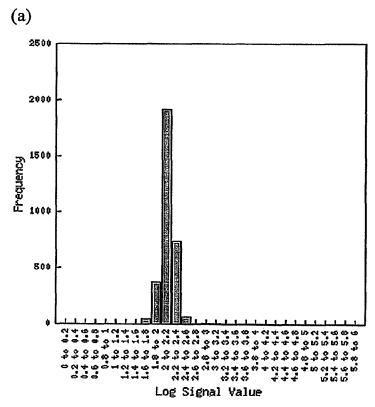


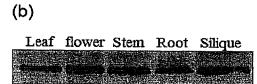


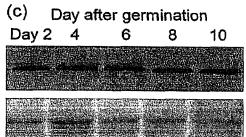
B

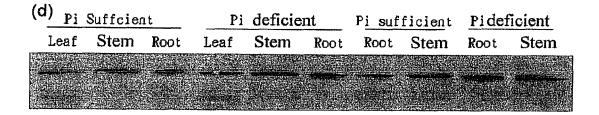


**FIG.** 7









**FIG. 8** 

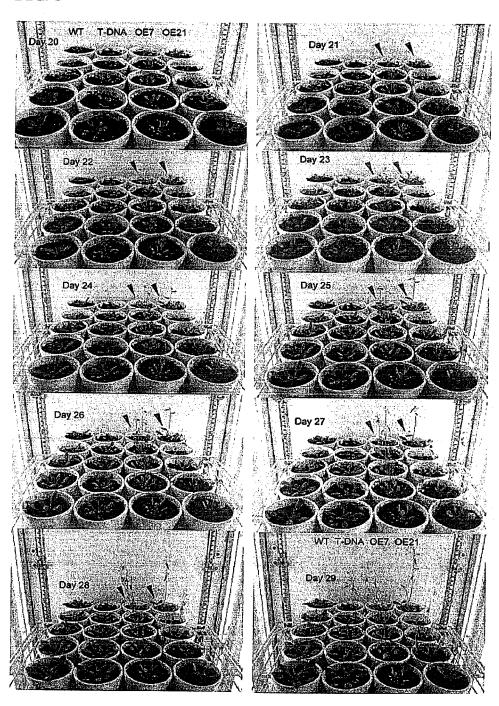
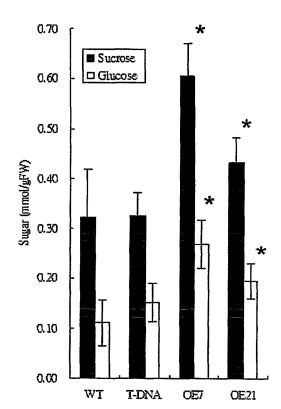


FIG. 9



\*Statistically (p<0.001) different from the WT (n = 10).

FIG. 10

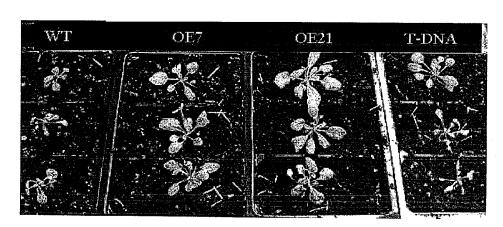
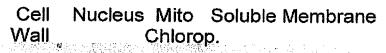


FIG. 11



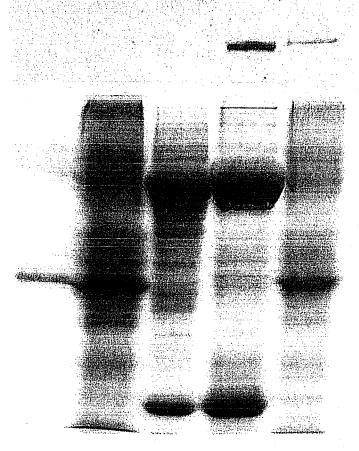


FIG. 12

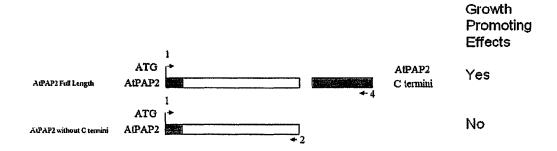
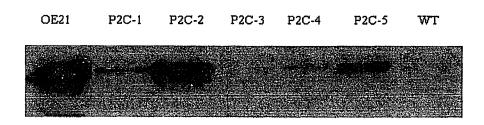


FIG. 13



#### METHOD FOR SPEEDING UP PLANT GROWTH AND IMPROVING YIELD BY INTRODUCING PHOSPHATASES IN TRANSGENIC PLANT

#### RELATED APPLICATIONS

This application is a divisional of U.S. application Ser. No. 12/640,674 filed Dec. 17, 2009, which claims priority to provisional application Ser. No. 61/138,918, filed on Dec. 18, 2008, the disclosures of which are incorporated herein by reference.

#### INCORPORATION OF SEQUENCE LISTING

A computer readable form of the sequence listing is contained in the file named "VRST002USD1\_ST25.txt" which is 143 kb (measured in MS-Windows) and was created on Jul. 9, 2013, which is filed herewith and herein incorporated by reference.

#### 1. TECHNICAL FIELD

The present disclosure provides methods that speeds up plant growth and elevates plant yields by introducing phosphatases with a C-terminal motif into plants. The present disclosure relates to phosphatases with a C-terminal motif, and their respectively encoded protein products, as well as fragments, derivatives, homologues, and variants thereof. Methods for introducing these genes into plants to (1) speed up the growth rate of plants, (2) to increase the sugar contents of plants, and (3) to increase of yield of plants, are provided.

#### 2. BACKGROUND

Purple acid phosphatases (PAPs) catalyze the hydrolysis of a wide range of activated phosphoric acid mono- and di-esters and anhydrides (Klabunde et al., 1996). The PAP proteins are characterized by seven conserved amino acid 40 residues (shown in bold face) in the five conserved motifs XDXX, XDXXY, GNH(D/E), XXXH, XHXH, which are involved in the coordination of the dimetal nuclear center (Fe³+-Me²+) in the active site (Li et al., 2002), where Me is a transition metal and Me²+ is mostly found to be Fe²+ in 45 mammalian, and Zn²+, or Mn²+ in plants (Klabunde and Krebs, 1997; Schenk et al., 1999).

Purple acid phosphatases are distinguished from the other phosphatases by their characteristic purple color, which is caused by a charge transfer transition at 560 nm from a 50 metal-coordinating tyrosine to the metal ligand Fe<sup>3+</sup> (Klabunde and Krebs, 1997; Schenk et al., 2000). Different from the other acid phosphatases, PAPs are insensitive to inhibition by tartrate, so they are also known as tartrateresistant acid phosphatases (TRAPs).

The biochemical properties of some plant PAPs have been characterized, firstly in red kidney bean, and later in soybean suspension cell, soybean seedlings, rice culture cells, spinach leaves, sweet potato tubers, tomato, yellow lupin seeds, *medicago* and *Arabidopsis*, etc. (Schenk et al., 1999). Plant 60 PAPs are generally considered to mediate phosphorus acquisition and redistribution based on their ability to hydrolyze phosphate compounds (Cashikar et al., 1997; Bozzo et al., 2004; Lung et al., 2008). Regulation of some plant PAPs transcripts by external phosphate level in medium or soil, 65 strongly suggest their involving in phosphate acquisition. For example, the transcription level of *Medicago* MtPAP1 in

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roots was increased under P stress, implicating a role in P acquisition or internal mobilization (Xiao et al., 2005; Xiao et al., 2006). Some plant PAPs could be secreted from root cells to extracellular environment, then hydrolyze various phosphate esters. Lung et al. purified a secreted PAP phosphatase from tobacco, which could hydrolyze broad substrates and help to alleviate P starvation (Lung et al., 2008). Certain plant PAPs can also hydrolyze phytate, a major storage compound of phosphorus in plants. Hegeman and Grabeu (2001) purified a novel PAPs (GmPhy) from the cotyledon of the germinating soybean seedlings. GmPhy was introduced into soybean tissue culture and was assayed to show phosphatase activity. Most recently, AtPAP15 and 23 in Arabidopsis sharing high sequence homology (73-15 52%) with this soybean PAP, were found to exhibit phytase activity (Zhu et al., 2005; Zhang et al., 2008).

Besides involvement in P acquisition, plant PAPs may perform some other physiological roles. For example, the PAPs AtACP5 (AtPAP17), SAP1, and SAP2 (del Pozo et al., 1999; Bozzo et al., 2002) display not only phosphatase but also peroxidase activity, suggesting their involvement in the removal of reactive oxygen compounds in plant organs. A pollen-specific PAP from Ester lily was suggested to function as an iron carrier in mature pollen (Kim and Gynheung, 1996). Other studies indicate that plant PAPs may also be involved in NaCl stress adaption or cell regeneration (Kaida, 2003; Liao et al., 2003).

In the *Arabidopsis* genome, twenty-nine potential PAP genes were identified based on sequence comparison.

Twenty-four of these putative enzymes contain seven conserved amino-acids residues involved in metal binding. One (AtPAP13) lacked four of these seven residues, and the other four (AtPAP14, 16, 28 and 29) lacked either the first, the second, or both motifs of the five conserved motifs. Twenty-eight are actively transcribed in *Arabidopsis* (Zhu et al., 2005)

To date, relatively little is known about AtPAPs biochemical properties and physiological roles, though several members have been characterized (del Pozo et al., 1999). AtPAP17 (AtACP5) was first known to be induced by phosphorus starvation. The transcription of AtPAP17 was also responsive to ABA, salt stress (NaCl), oxidative stress (H<sub>2</sub>O<sub>2</sub>) and leaves senescence, according to GUS activity assay. No alteration in the expression of AtPAP17 was observed during the nitrogen or potassium starvation, and paraquat or salicylic acid. Like the other type 5 acid phosphatases, AtPAP17 displayed peroxidation activity, which may be involved in the metabolism of reactive oxygen species in stressed or senescent parts of plants.

Besides AtPAP17, several AtPAPs were found to be involved in phosphorus metabolism in *Arabidopsis*. Root secretion of AtPAP12 was induced by P stress, and its regulation was mainly at transcriptional level (Patel et al., 1998; Coello, 2002/11). AtPAP4, as well as AtPAP10, 55 AtPAP11 and AtPAP12 were involved in phosphorus starvation response since their transcription levels increased during phosphate deprivation (Li et al., 2002; Wu et al., 2003). In contrast, AtPAP20, 21 and 22 were irrespective to P starvation and expressed constitutively in Pi sufficient or deficient condition. Fluorescent signals were detected in the cytoplasm via the baculovirus expression system, indicating that they may function in the cytoplasm (Li and Wang, 2003).

AtPAP26 was purified and characterized from Pi-starved *Arabidopsis* suspension cell culture (Veljanovski et al., 2006). It exists as a homodimer with 55 kDa glycosylated protein, showing wide substrate specificity with the highest

activity against phosphoenolpyruvate (PEP) and polypeptide phosphate. AtPAP26 also displayed alkaline peroxidase activity with the probable roles in the metabolism of reactive oxygen species. Proteomic study suggested that it may be localized in vacuole, and involved in recycling Pi from 5 intracellular P metabolites (Shimaoka et al., 2004).

PAPs can act on a wide range of substrates, but not all of them exhibit phytase activity. An enzyme assay involving the GST-AtPAP23 fusion protein revealed that AtPAP23 exhibits phytase activity. A GUS study showed that 10 AtPAP23 is exclusively expressed in the flower of the Arabidopsis, and may play certain roles in flower development (Zhu et al., 2005). In a recent report, a recombinant AtPAP15 expressed and partial purified in E. coli and yeast was also found to exhibit phytase activity) (Zhang et al., 15 2008). It was proposed that AtPAP15 may be involved in ascorbic acid biosynthesis with the end product myo-inositol of phytate hydrolysis as the precursor of ascorbic acid

As stated above, most of the functions of characterized 20 plant PAPs are related to phosphorus metabolism. None of the functionally or biochemically characterized plant PAPs carry transmembrane motif, and none of them were shown to be associated with membrane. Furthermore, to date, no AtPAPs or any plant PAPs, have been showed to affect sugar 25 signalling and carbon metabolism in plant.

The first report of transgenic expression of plant PAP in plant was reported in 2005 (Xiao et al., 2005). The PAPphosphatase gene from Medicago (MtPHY1) was expressed in transgenic Arabidopsis, resulting in increased capacity of 30 Pacquisition from phytate in agar culture (Xiao et al., 2005). Nonetheless, the growth performance of the plants was not reported to be different under normal growth.

#### 3. SUMMARY

The present disclosure provides a method that speeds up plant growth and elevates plant yields by introducing phosphatases with a C-terminal motif into plants, Phosphatases with a C-terminal motif, and their respectively encoded 40 protein products, as well as fragments, derivatives, homologues, and variants thereof are disclosed. Methods for introducing this class of genes into plants to speed up the growth rate of plants, to increase the sugar contents of plants, and to increase of yield of plants, are provided. 45 Without wishing to be bound by any particular theory, the C-terminal motif is believed to function as a transmembrane structural element (transmembrane motif).

As stated above in the Background section, most of the functions of characterized plant PAPs are related to phos- 50 phorus metabolism. None of the functionally or biochemically characterized plant PAPs carry transmembrane motif, and none of them were shown to be associated with membrane. Furthermore, to date, no AtPAPs or any plant PAPs, have been showed to affect sugar signalling and carbon 55 nant expression vectors, comprising a nucleic acid molecule metabolism in plant.

The first report of transgenic expression of plant PAP in plant was reported in 2005 (Xiao et al., 2005). The PAPphosphatase gene from Medicago (MtPHY1) was expressed in transgenic Arabidopsis, resulting in increased capacity of 60 P acquisition from phytate in agar culture. Nonetheless, the growth performance of the plants was not reported to be different under normal growth.

We also produced transgenic tobacco and Arabidopsis that overexpressed AtPAP15, a PAP with phosphatase activ- 65 ity, which does not carry any C-terminal motif equivalent to that of AtPAP2; phosphatase activity was secreted into

extracellular growth medium. Significant secretion of phosphatase activity was observed in the transgenic plants and the transgenic plants showed larger biomass than the control plants in agar and soil supplemented with exogenous phytate. Higher P content was also obtained in overexpressed transgenic lines in phytate treatment. However, the growth of transgenic plants overexpressing AtPAP15 did not show any difference in growth phenotypes when it was compared with the wild-type, under treatments of K—P or No-P, or in soil.

Here, we have developed a technology to speed up plant growth and improve seed yield by overexpressing a phosphatase with a C-terminal motif in plants. An example is the use of a purple acid phosphatase (PAP). This disclosure is the first report to show that overexpressing a phosphatase with a C-terminal motif in transgenic plant is able to speed up the growth of the plants, to increase the sugar contents of plants, and to increase the yield of plants, by altering the carbon metabolism of the plants.

The present advances are based, in part, on the characterization of a group of purple acid phosphatases (SEQ ID NOS: 1-8 and 18-47) from plants and the observations that overexpression of a purple acid phosphatase (AtPAP2, SEQ ID NO:1) of this group in plants resulted in rapid plant growth, higher sugar content, and higher yield. Accordingly, nucleotide sequences of a group of purple acid phosphatase genes (SEQ ID NOs:1, 3, 5, 7, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44 and 46), which share a C-terminal motif/domain, from plants and amino acid sequences of their encoded proteins (SEQ ID NOS:2, 4, 6, 8, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45 and 47), as well as fragments, derivatives, homologues, and variants thereof, as defined herein, are disclosed. Furthermore, nucleic acid molecules encoding the polypeptides of interest, and include 35 cDNA, genomic DNA, and RNA, are disclosed.

As used herein, italicizing the name of a gene shall indicate the gene, in contrast to its encoded protein or polypeptide product which is indicated by the name of the gene in the absence of any italicizing. For example, "Gene" shall mean the Gene gene, whereas "Gene" shall indicate the protein or polypeptide product of the Gene gene.

In one embodiment, isolated nucleic acid molecules hybridize under stringent conditions, as defined herein, to nucleic acids having the sequence of SEQ ID NOS: 1, 3, 5, 7, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44 or 46, or homologues thereof, wherein the nucleic acid molecules encode proteins or polypeptides which exhibit at least one structural and/or functional feature of the polypeptides of the invention.

Another embodiment includes, nucleic acid molecules, which are suitable for use as primers or hybridization probes for the detection of nucleic acids encoding one of the disclosed phosphatase polypeptides or other sequences.

Yet another embodiment includes vectors, e.g., recombiof the invention. Furthermore, host cells containing such a vector or engineered to contain and/or express a nucleic acid molecule of the invention and host cells containing a nucleotide sequence of the invention operably linked to a heterologous promoter are disclosed.

A further embodiment includes methods for preparing a polypeptide of the invention by a recombinant DNA technology in which the host cells containing a recombinant expression vector encoding a polypeptide of the invention or a nucleotide sequence encoding a polypeptide of the invention operably linked to a heterologous promoter, are cultured, and the polypeptide of the invention are produced.

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In still further another embodiment, a transgenic plant contains a nucleic acid molecule which encodes an isolated polypeptides or proteins comprising the five conserved motifs of purple acid phosphatases, including XDXX, XDXXY, GNH(D/E), XXXH, XHXH, and linked to a 5 C-terminal motif.

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Embodiments further provide antibodies that immunospecifically bind a polypeptide of the invention. Such antibodies include, but are not limited to, antibodies from various animals, humanized, chimeric, polyclonal, monoclonal, bispecific, multi-specific, single chain antibodies, Fab fragments, F(ab')<sub>2</sub> fragments, disulfide-linked Fvs, fragments containing either a VL or VH domain or even a complementary determining region (CDR), that immunospecifically binds to a polypeptide of the invention.

In an additional embodiment, method for detecting the presence, activity or expression of a polypeptide of the invention or similar polypeptide in a biological material, such as cells, culture media, and so forth are provided. The increased or decreased activity or expression of the polypeptide in a sample relative to a control sample can be determined by contacting the biological material with an agent that can detect directly or indirectly the presence, activity or expression of the polypeptide of the invention. In a particular embodiment, such an agent is an antibody or a 25 fragment thereof which immunospecifically binds to a one of the disclosed polypeptides.

In a still another embodiment, a fusion protein comprising a bioactive molecule and one or more domains of a disclosed polypeptide or fragment thereof is provided. In particular, <sup>30</sup> fusion proteins comprising a bioactive molecule recombinantly fused or chemically conjugated (including both covalent and non-covalent conjugations) to one or more domains of a disclosed polypeptide or fragments thereof.

We also produced transgenic tobacco and *Arabidopsis* 35 that overexpressed AtPAP15, a PAP with phosphatase activity, which does not carry any C-terminal motif and was found to be secreted into extracellular growth medium. Significant secretion of phosphatase activity was observed in the transgenic plants and the transgenic plants showed larger 40 biomass than the control plants in agar and soil supplemented with exogenous phytate. Higher P content was also obtained in overexpressed transgenic lines in phytate treatment. However, the growth of transgenic plants overexpressing AtPAP15 did not show any difference in growth 45 phenotypes when it was compared with the wild-type, under treatments of K—P or No P, or in soil.

In conclusion, this disclosure is the first report to show that overexpressing a phosphatase with a C-terminal motif in transgenic plant is able to speed up the growth of the plants, 50 to increase the sugar contents of plants, and to increase the yield of plants, by altering the carbon metabolism of the plants.

#### 3.1 DEFINITIONS

The term "acidic" or "acid pH" as used herein refers to a pH value of less than about 6.0.

The term "homologue" as used herein refers to a polypeptide that possesses a similar or identical function to 60 polypeptides encoded by SEQ ID NOS: 2, 4, 6, 8, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45 and 47, and/or a fragment of these polypeptides, that do not have an identical amino acid sequence of these polypeptides and/or a fragment of these polypeptides. A polypeptide that has a similar 65 amino acid sequence included in the definition of the term "homologue" includes a polypeptide that satisfied at least

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one of the following: (i) polypeptide having an amino acid sequence that is one or more of at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 90%, and at least about 98% identical. (ii) a polypeptide encoded by a nucleotide sequence that is one or more of at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 90%, and at least about 98% identical and/or conservatively substituted to one or more of the nucleotide sequences encoding the polypeptides of SEQ ID NOS: 2, 4, 6, 8, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45 and 47, and/or a fragment of the these polypeptides; (iii) a polypeptide encoded by a nucleotide sequence that hybridizes under stringent conditions as defined herein to one or more of nucleotide sequences SEQ ID NOS: 1, 3, 5, 7, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44 and 6; (iv) a polypeptide having an amino acid sequence that is one or more of at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 90, and at least about 98% identical and/or conservatively substituted; (v) a nucleic acid sequence encoding an amino acid sequence that is one or more of at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 90%, and at least about 98% identical and/or conservatively substituted; (vi) a fragment of any of the polypeptides or nucleic acid sequences described in (i) through (v) having one of at least 20 amino acid residues, at least 25 amino acid residues, at least 40 amino acid residues, at least 80 amino acid residues, at least 90 amino acid residues, at least 100 amino acid residues, at least 125 amino acid residues, at least 150 amino acid residues, at least 175 amino acid residues, at least 200 amino acid residues, at least 225 amino acid residues, at least 250 amino acid residues, at least 275 amino acid residues, at least 300 amino acid residues, at least 325 amino acid residues, at least 350 amino acid residues, or at least 375 amino acid residues; (vii) a polypeptide with similar structure and function or a nucleotide sequence encoding a polypeptide with similar structure and function, exhibiting the antigenicity, immunogenicity, catalytic activity, and other readily assayable activities, to polypeptides encoded by SEQ ID NOS: 2, 4, 6, 8, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45 and 47, and/or a fragment of these polypeptides, refers to a polypeptide that has a similar secondary, tertiary, or quaternary structure of these polypeptides, or a fragment of these polypeptides. The structure of a polypeptide can be determined by methods known to those skilled in the art, including but not limited to, X-ray crystallography, nuclear magnetic resonance, and crystallographic electron microscopy. The term "homologue" is used herein to describe a sequence that has sequence homology. A sequence having sequence homology can be made using standard molecular biology techniques including site-di-55 rected mutagenesis including insertion or deletion of sequences. The term "homologue" is not limited to homologous genes or proteins originating from different species and expressly includes artificial modification to the sequences disclosed herein.

The term "conservatively substituted variant" refers to a polypeptide or a nucleic acid sequence encoding a homologue polypeptide in which one or more amino acid residues or codons have been modified by conservative substitution with an amino acid residue or a codon coding for an amino acid residue of similar chemical-type, as described below.

The term "an antibody or an antibody fragment which immunospecifically binds to polypeptides encoded by SEQ

ID NOS: 2, 4, 6, 8, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45 and 47," as used herein refers to an antibody or a fragment thereof that immunospecifically binds to polypeptides encoded by SEQ ID NOS: 2, 4, 6, 8, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45 and 47, or a fragment 5 of these polypeptide and does not non-specifically bind to other polypeptides. An antibody or a fragment thereof that immunospecifically binds to polypeptides encoded by SEQ ID NOS: 2, 4, 6, 8, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45 and 47, or a fragment of these polypeptide, may 10 cross-react with other antigens. Preferably, an antibody or a fragment thereof that immunospecifically binds to polypeptides encoded by SEQ ID NOS: 2, 4, 6, 8, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45 and 47, or a fragment of these polypeptides, does not cross-react with other antigens. 15 An antibody or a fragment thereof that immunospecifically binds to polypeptides encoded by SEQ ID NOS: 2, 4, 6, 8, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45 and 47, or a fragment of these polypeptide, can be identified by, for example, immunoassays or other techniques known to those 20 skilled in the art. An antibody or an antibody fragment which immunospecifically binds polypeptides encoded by SEQ ID NOS: 2, 4, 6, 8, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45 and 47, may be interchangeably referred to as "anti-PAP antibody".

The term "derivative" as used herein refers to a given peptide or protein that is otherwise modified, e.g., by covalent attachment of any type of molecule, preferably having bioactivity, to the peptide or protein, including the incorporation of non-naturally occurring amino acids. The resulting bioactivity retains one or more biological activities of the peptide protein.

The term "fragment" as used herein refers to a fragment of a nucleic acid molecule containing one of at least about 25, at least about 30, at least about 35, at least about 40, at 35 least about 45, at least about 100, at least about 150, at least about 200, at least about 250, at least about 300, at least about 350, at least about 400, at least about 450, at least about 500, at least about 550, at least about 600, at least about 650, at least about 700, at least about 750, at least 40 about 800, at least about 850, at least about 900, at least about 950, at least about 1000, at least about 1050, at least about 1100, at least about 1150, at least about 1200, at least about 1250, at least about 1300, at least about 1350, from about 500 to about 2000, from about 1000 to about 2000 45 from about 200 to about 500, from about 500 to about 1000, form about 1000 to about 1500, and from about 1500 to about 2000 nucleic acid bases in length of the relevant nucleic acid molecule and having at least one functional feature of the nucleic acid molecule (or the encoded protein 50 has one functional feature of the protein encoded by the nucleic acid molecule); or a fragment of a protein or a polypeptide containing one or more of at least about 5, at least about 10, at least about 15, at least about 20, at least about 25, at least about 30, at least about 35, at least about 55 40, at least about 45, at least about 50, at least about 55, at least about 60, at least about 65, at least about 70, at least about 75, at least about 80, at least about 90, at least about 100, at least about 120, at least about 140, at least about 160, at least about 180, at least about 200, at least about 220, at 60 least about 240, at least about 260, at least about 280, at least about 300, at least about 320, at least about 340, at least about 360, from about 250 to about 660, from about 350 to about 660, form about 450 to about 660, and form about 550 to about 660 amino acid residues in length of the relevant 65 protein or polypeptide and having at least one functional feature of the protein or polypeptide, such functional fea8

tures include ability to bind a Fe<sup>3+</sup>-Me<sup>2+</sup> dimetal nuclear center and form a C-terminal motif.

An "isolated" nucleic acid molecule is one which is separated from other nucleic acid molecules which are present in the natural source of the nucleic acid molecule. Moreover, an "isolated" nucleic acid molecule, such as a cDNA molecule, can be substantially free of other cellular materials, or culture medium when produced by recombinant techniques, or substantially free of chemical precursors or other chemicals when chemically synthesized, but excludes nucleic acid molecules present in recombinant DNA libraries. In a preferred embodiment, nucleic acid molecules encoding the disclosed polypeptides/proteins are isolated or purified.

The term "operably linked" as used herein refers to when transcription under the control of the "operably linked" promoter produces a functional messenger RNA, translation of which results in the production of the polypeptide encoded by the DNA operably linked to the promoter.

The term "under stringent condition" refers to hybridization and washing conditions under which nucleotide sequences having homology to each other remain hybridized to each other. Such hybridization conditions are described 25 in, for example but not limited to, Current Protocols in Molecular Biology, John Wiley & Sons, N.Y. (1989), 6.3.1-6.3.6; Basic Methods in Molecular Biology, Elsevier Science Publishing Co., Inc., N.Y. (1986), pp. 75-78, and 84-87; and Molecular Cloning, Cold Spring Harbor Laboratory, N.Y. (1982), pp. 387-389, and are well known to those skilled in the art. A preferred, non-limiting example of stringent hybridization conditions is hybridization in 6× sodium chloride/sodium citrate (SSC), 0.5% SDS at about 68° C. followed by one or more washes in 2×SSC, 0.5% SDS at room temperature. Another preferred, non-limiting example of stringent hybridization conditions is hybridization in 6×SSC at about 45° C. followed by one or more washes in 0.2×SSC, 0.1% SDS at about 50-65° C.

The term "variant" as used herein refers either to a naturally occurring allelic variation of a given peptide or a recombinantly prepared variation of a given peptide or protein in which one or more amino acid residues have been modified by amino acid substitution, addition, or deletion.

The term "aligned" as used herein refers to a homology alignment between two or more sequences using a standard algorithm such as BLAST (http://blast.ncbi.nlm.nih.gov/Blast.cgi).

The term "predicted to form a transmembrane motif by TMHMM analysis" or "predicted to form a C-terminal motif by TMHMM analysis" (http://www.cbs.dtu.dk/services/TMHMM/) herein refers to a probability that is equal to or greater than about 0.5.

#### BRIEF DESCRIPTION OF THE FIGURES

The following figures illustrate the embodiments and are not meant to limit the scope of the invention encompassed by the claims.

FIG. 1 shows the phylogenetic tree of PAP-like sequences in the *Arabidopsis* genome. Twenty-nine PAPs were aligned using ClustalX and the phylogenetic tree was created by the neighbor-joining algorithm of the MEGA4 program. The accession numbers of the PAP-like, transmembrane-like C-terminal motif containing, polypeptide from *Zea mays* (ZmPAP2) and *Oryza sativa* (OsPAP2) were ACG47621 and BAC15853.1, respectively.

FIG. 2A is the amino acid alignment of the C-terminal transmembrane-like motifs in AtPAP2 (SEQ ID NO. 2) with other PAP sequences.

FIG. 2B is the amino acid alignment of AtPAP2 (SEQ ID NO. 2) with other PAP sequences, showing the full length of 5 each sequence. These sequences include homologous sequences from B. napus (BnPAP2) SEQ ID NO. 47, G. max (GmPAP2, SEQ ID NO. 6) and Z. may (ZmPAP2, SEQ ID NO. 8). The five conserved motifs (XDXX, XDXXY, GNH (D/E), XXXH, XHXH) are boxed. Residues in shades have 10 low or no homology. Hydrophobic motifs at the C-termini of these polypeptides are underlined by a bar (614th-636th amino acid), which is absent from the sequence of AtPAP15. As shown, AtPAP15 (SEQ ID NO:67) does not have a C-terminal region corresponding to the other PAP 15 sequences.

FIG. 3 shows that a unique hydrophobic motif is present at the C-termini of AtPAP2 and ZmPAP2 by TMHMM analysis. This transmembrane-like C-terminal motif is absent from AtPAP15.

FIG. 4 shows the characteristics of the T-DNA lines. The T-DNA line (Salk\_013567) was obtained from TAIR. The AtPAP2 genomic sequence carries two exons and the T-DNA was inserted in exon 2 and causes a disruption of the AtPAP2 mRNA (a). Three PCR primers (A, B and C) were 25 designed for the differentiation of the wild-type (WT) and the T-DNA line (atpap2-8) and they were used for PCR screening of genomic DNA extracted from WT and the T-DNA line (b). Total RNA was extracted from 10-day-old seedlings grown on MS with 2% sucrose using the TRIzol 30 RNA isolation method and were used for RT-PCR (c). 50 µg of seedlings proteins were loaded for Western blotting studies, using the anti-AtPAP2 specific antiserum (Section

FIG. 5 is the schematic diagram of the expression vector 35 pBV-AtPAP2. CaMV 35S:35S promoter of the cauliflower mosaic virus; NOS: polyadenylation signal of nopaline synthase gene; aadA: bacterial streptomycin/spectinomycin resistance gene encoding aminoglycoside-3"-adenyltransferase; pNOS:BAR: bialaphos resistance gene under the 40 control of the nopaline synthase promoter; born: basis of mobility from pBR322; ColE1: replication origin from pBR322; pVS1-REP: replication origin from pVS1; pVS1-STA: STA region from pVS1 plasmid; LB: left border wicz et al., 1994).

FIG. 6A shows the results of the Western blot analysis of the overexpression lines (OE), wild-type (WT), T-DNA and the complementation lines (CP) of AtPAP2 and FIG. 6B shows the results of the Western blot analysis of the over- 50 expression lines (C-15) and wild-type (WT) of AtPAP15.

FIG. 7 shows the expression analysis of AtPAP2. The mRNA expression profile was analysed by the Spot History program of NASC (a). The protein expression profiles of 30 day old, soil-grown plant (b), seedlings germinated on MS 55 agar (c) and 2 week old plants transferred to Pi-sufficient/ Pi-deficient MS agar for 3 days (d), were analyzed by Western blotting using the anti-PAP2 antiserum.

FIG. 8 shows the growth performance of the wild-type, T-DNA and overexpression lines in soil. Seeds were germi- 60 nated in MS agar with 2% sucrose for 10 days. Seedlings with 2 small visible rosette leaves (~1 mm) were transferred to soil and grown under 16 h/8 h light/dark cycles.

FIG. 9 shows the levels of sucrose and glucose in the rosette leaves of 21-day-old, soil grown seedlings.

FIG. 10 shows the recovery of various lines after prolonged darkness treatment. Seeds were germinated in MS 10

agar with 2% sucrose for 10 days. Seedlings with 2 small visible rosette leaves (~1 mm) were transferred to soil and grown for 12 days under 16 h/8 h light/dark cycles. The lights of the growth chamber were then switched off for 12 days and the plants were allowed to recover under 16 h/8 h light/dark cycles for 1 week. n=9-12 per line.

FIG. 11. Detection of AtPAP2 protein in subcellular fractions by Western blotting. Mito.: Mitochondria; Chlorop.: Chloroplasts.

FIG. 12. shows a schematic representation of two vector constructs incorporating the AtPAP2 gene.

FIG. 13. shows Western blot analysis results for overexpression of AtPAP2 proteins missing the C-terminal motif.

#### DETAILED DESCRIPTION

5.1 Method of Speeding Up Plant Growth and Improving Crop Yield

The present disclosure provides a method that speeds up plant growth and elevates plant yields by introducing phosphatases with a C-terminal motif into plants. In an embodiment, the present disclosure relates to a class of genes of purple acid phosphates, and their respectively encoded protein products, as well as fragments, derivatives, homologues, and variants thereof. Methods for introducing this class of genes into plants to speed up the growth rate of plants, to increase the sugar contents of plants, and to increase of yield of plants, are provided.

A group of purple acid phosphatases (PAPs) which carry seven conserved amino acid residues (shown in bold face) in the five conserved motifs XDXX (example GDXG (SEQ ID NO: 48)), XDXXY (SEQ ID NO: 49), GNH(D/E) (SEQ ID NOS: 50-51), XXXH (example ZXGH (SEQ ID NO: 52)), XHXH (SEQ ID NO: 53), where X is any amino acid and Z is any amino acid selected from L, I, V, F, and M, and a transmembrane-like motif at their C-termini were identified in the genomes of a number of plants (FIGS. 1, 2A, and 2B). The presence of the C-terminal transmembrane-like motif enables the localization of this group of PAN to the membrane fraction (FIGS. 3 and 11). This property makes this group of PAPs differ from the other previously characterized PAPs because all previously characterized PAPs did not T-DNA repeat; RB: right border T-DNA repeat. (Hajdukie- 45 carry any C-terminal motif (FIGS. 2A, 2B, and 3). By using the protein sequence of a representative gene of this group, AtPAP2, to blast the NCBI database and various EST databases, a number of genomic or cDNA sequences were identified to encodes polypeptides that carry the five conserved motifs XDXX, XDXXY, GNH(D/E), XXXH, XHXH of PAPs and a transmembrane motif at their C-termini (FIG. 2B).

> The introduction of a representative gene of this group of phosphatases, AtPAP2, into the genome of Arabidopsis by transgenic technology produced transgenic Arabidopsis that grew faster than the wild-type plants (FIG. 8), and the yield of seeds were elevated by approximately 40% (Table 3). However, transgenic plant that expressed AtPAP15 did not show these phenotypes. The sugar contents, including glucose and sucrose, in the leaf of the transgenic lines, were also found to be higher than that of the wild-types (FIG. 9).

Thus, this disclosure provides a method that speeds up plant growth and elevates plant yields by introducing phosphatases into plants. In an embodiment, a group of genes of purple acid phosphatases, and their respectively encoded protein products, as well as fragments, derivatives, homologues, and variants thereof are described.

5.2 Homologues, Derivatives, and Variants of Phosphatases

In addition to the nucleic acid molecules (SEQ ID NOS: 1, 3, 5, 7, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44 and 46) and polypeptides (SEQ ID NOS: 2, 4, 8, 19, 21, 523, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45 and 47) described in claims 9-16, the nucleic acid molecules and polypeptides also encompass those nucleic acid molecules and polypeptides having a common biological activity, similar or identical structural domain and/or having sufficient nucleotide sequence or amino acid identity (homologues) to those of the nucleic acid molecules and polypeptides described above.

Such common biological activities of the polypeptides include antigenicity, immunogenicity, catalytic activity especially phosphatase activity, ability to bind a Fe³+-Me²+ dimetal nuclear center, fold into or form a transmembrane-like C-terminal motif and other activities readily assayable by the skilled artisan.

A polypeptide that has a similar amino acid sequence 20 (homologue) refers to a polypeptide that satisfied at least one of the following: (i) a polypeptide having an amino acid sequence that is one of at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 90%, and at least 25 about 95%, and at least about 98% identical and/or conservatively substituted to the amino acid sequence of a AtPAP2 (SEQ ID NO: 2) and/or other PAPs with a transmembranelike C-terminal motifincluding SEQ ID NOS: 4, 6, 8, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45 and/or 47, a 30 fragment of AtPAP2, and having at least one biological feature of the described polypeptides; (ii) a polypeptide encoded by a nucleotide sequence that is one of at least about 30%, at least about 40%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least 35 about 80%, at least about 90%, at least about 95%, and at least about 98% identical to the nucleotide sequence encoding AtPAP2 (SEQ ID NO: 1) and/or other PAPs with a transmembrane-like C-terminal motif including SEQ ID NOS: 3, 5, 7, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 40 42, 44 and/or 46, a fragment of AtPAP2 and having at least one structural and/or biological feature of AtPAP2; (iii) a polypeptide encoded by a nucleotide sequence that hybridizes under stringent conditions as defined herein to a nucleotide sequence encoding AtPAP2 (SEQ ID NO: 1) and/or 45 other PAPs with a motif including SEQ ID NOS: 3, 5, 7, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44 and/or 46, a fragment of AtPAP2 and having at least one structural and/or biological feature of AtPAP2. A polypeptide with similar structure to AtPAP2, or a fragment of AtPAP2, refers 50 to a polypeptide that has a similar secondary, tertiary, or quaternary structure of AtPAP2, a fragment of AtPAP2 and has at least one functional feature of a AtPAP2, including one or more of ability to bind a Fe<sup>3+</sup>-Me<sup>2+</sup> dimetal nuclear center and fold into or form a transmembrane-like C-termi- 55 nal motif. The structure of a polypeptide can be determined by methods known to those skilled in the art, including but not limited to, X-ray crystallography, nuclear magnetic resonance, and crystallographic electron microscopy.

Those having skill in the art will readily recognized that 60 mutations, deletions or insertions can be made in any of the sequences disclosed herein, including SEQ ID NOS: 1-8 and 18-47, without affecting function. Sequences useful in practicing the embodiments include sequences having homology to SEQ ID NOS: 1-8 and 18-47 and being a protein, 65 polypeptide, or polynucleotide coding for such protein or peptide having functionality to bind a dimetal nuclear center

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(Fe<sup>3+</sup>-Me<sup>2+</sup>) and being a protein, polypeptide, or polynucleotide coding for such protein or peptide having a C-terminal motif. That is, those skilled in the art will recognize that many mutations can be made to any of SEQ ID NOS: 1-8 and 18-47 without affecting the catalytic functionality nor interrupting the transmembrane-like C-terminal motif. Such modified sequences that maintain catalytic activity and a transmembrane-like C-terminal motif are defined as homologues to SEQ ID NOS: 1-8 and 18-47 and are including within the scope of useful sequences.

In one embodiment, such homologues can have about 30% or more identity to the sequences disclosed herein. In another embodiment, such homologues can have about 40%or more identity to the sequences disclosed herein. In yet another embodiment, such homologues can have about 50% or more identity to the sequences disclosed herein. In sill yet another embodiment, such homologues can have about 60% or more identity to the sequences disclosed herein. In even sill yet another embodiment, such homologues can have about 70% or more identity to the sequences disclosed herein. In a further embodiment, such homologues can have about 80% or more identity to the sequences disclosed herein. In yet a still further embodiment, homologues can have about 90% or more identity to the sequences disclosed herein. In a still further embodiment, homologues can have about 98% or more identity to the sequences disclosed herein.

Those having skill in the art will recognize that mutations can be made to proteins and peptides and/or to polynucleotides coding for protein and peptides or complementary thereto that substitute amino acid residue for other amino acids residues having similar chemical properties (conservative substitutions) and that such mutations are less likely to cause structural changes that affect functionality including catalytic activity and/or the function of a transmembranelike C-terminal motif. Conservatively substituting amino acids are substituting an amino acid residue belong to any of the following 11 chemical groups with another amino acid from the same chemical group: (1) acidic (negatively charged) amino acids such as aspartic acid and glutamic acid; (2) basic (positively charged) amino acids such as arginine, histidine, and lysine; (3) neutral polar amino acids such as glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine; (4) neutral nonpolar (hydrophobic) amino acids such as alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan, and methionine; (5) amino acids having aliphatic side chains such as glycine, alanine, valine, leucine, and isoleucine; (6) amino acids having aliphatic-hydroxyl side chains such as serine and threonine; (7) amino acids having amide-containing side chains such as asparagine and glutamine; (8) amino acids having aromatic side chains such as phenylalanine, tyrosine, and tryptophan; (9) amino acids having basic side chains such as lysine, arginine, and histidine; (10) amino acids having sulfur-containing side chains such as cysteine and methionine; (11); amino acids having similar geometry and hydrogen bonding patterns such as aspartic acid, asparagine, glutamic acid and glutamine.

In one embodiment, homologues can have about 30% or more identity and/or conservative substitutions to the sequences disclosed herein. In another embodiment, homologues can have about 40% or more identity and/or conservative substitutions to the sequences disclosed herein. In yet another embodiment, homologues can have about 50% or more identity and/or conservative substitutions to the sequences disclosed herein. In still yet another embodiment, homologues can have about 60% or more identity and/or

conservative substitutions to the sequences disclosed herein. In a further embodiment, homologues can have about 70% or more identity and/or conservative substitutions to the sequences disclosed herein. In a still further embodiment, homologues can have about 80% or more identity and/or 5 conservative substitutions to the sequences disclosed herein. In still another embodiment, homologues can have about 90% or more identity and/or conservative substitutions to the sequences disclosed herein. In still another further embodiment, homologues can have about 98% or more 10 identity and/or conservative substitutions to the sequences disclosed herein.

Embodiments further provide isolated nucleic acid molecules which comprise or consist of one or more of at least about 25, at least about 30, at least about 35, at least about 15 40, at least about 45, at least about 100, at least about 150, at least about 200, at least about 250, at least about 300, at least about 350, at least about 400, at least about 450, at least about 500, at least about 550, at least about 600, at least about 650, at least about 700, at least about 750, at least 20 about 800, at least about 850, at least about 900, at least about 950, at least about 1000, at least about 1050, at least about 1100, at least about 1150, at least about 1200, at least about 1250, at least about 1300, at least about 1350, from from about 200 to about 500, from about 500 to about 1000, form about 1000 to about 1500, and from about 1500 to about 2000 nucleotides of the nucleotide sequences of SEQ ID NOS: 1, 3, 5, 7, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44 and 46, or a complement thereof encoding a 30 protein or polypeptide having one or more activity of the amino acid sequences of their encoded proteins (SEQ ID NOS: 2, 4, 6, 8, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45 and 47). The activity includes one or more of antigenicity, immunogenicity, catalytic activity (e.g., phos- 35 phatase activity), ability to bind a Fe<sup>3+</sup>-Me<sup>2+</sup> dimetal nuclear center, fold into or form a transmembrane-like C-terminal motif, and other activities readily assayable.

Embodiments provide isolated polypeptides or proteins consisting of an amino acid sequence that contains one of 40 about 5, at least about 10, at least about 15, at least about 20, at least about 25, at least about 30, at least about 35, at least about 40, at least about 45, at least about 50, at least about 55, at least about 60, at least about 65, at least about 70, at least about 75, at least about 80, at least about 90, at least 45 about 100, at least about 120, at least about 140, at least about 160, at least about 180, at least about 200, at least about 220, at least about 240, at least about 260, at least about 280, at least about 300, at least about 320, at least from about 350 to about 660, form about 450 to about 660, and form about 550 to about 660 amino acid bases in length of the relevant protein or polypeptide and having at least one functional feature of the protein or polypeptide, such functional features including ability to bind a Fe<sup>3+</sup>-Me<sup>2+</sup> dimetal 55 nuclear center and form a transmembrane-like C-terminal

Additional embodiments are any of the phosphatases and homologues thereof with the identity and/or conservative substitutions to SEQ ID NOS: 1-8 and 18-47 described 60 above that additionally consist of a protein, polypeptide, or polynucleotide encoding a protein having the five conserved motifs in purple acid phosphatases, including XDXX, XDXXY, GNH(D/E), XXXH, XHXH, where X is any amino acid. In one embodiment, the described phosphatases 65 and homologues thereof consist of a protein, polypeptide, or polynucleotide encoding one of the sequences YHVCIGN-

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HEYDF (SEQ ID NO: 54) and YHVCIGNHEYDW (SEQ ID NO: 55). In one embodiment, the described phosphatases and homologues thereof consist of a protein, polypeptide, or polynucleotide encoding a sequence of amino acid residues having one of the sequences YHVCIGNHEYD(W/F) (SEO ID NO: 54) and YHVCIGNHEYN(W/F) (SEO ID NO: 55) or a protein, polypeptide, or polynucleotide encoding a homologue to one of the foregoing sequences with only conservative substitutions, as described above, to those sequences. In yet another embodiment, the described phosphatases and homologues thereof consist of a protein, polypeptide, or polynucleotide encoding a sequence of amino acid residues having one of the sequences GNHE (SEQ ID NO: 51) and GNHD (SEQ ID NO: 50). In still yet another embodiment, the described phosphatases and homologues thereof consist of a protein, polypeptide, or polynucleotide encoding a sequence of amino acid residues having one of the sequences GNHE (SEQ ID NO: 51) and GNHD (SEQ ID NO: 50) or a protein, polypeptide, or polynucleotide encoding a protein having a homologous sequence to one of the foregoing sequences SEQ 1N NOS: 50-51 with only conservative substitutions.

Additional embodiments are any of the phosphatases and about 500 to about 2000, from about 1000 to about 2000, 25 homologues thereof with the identity and/or conservative substitutions to SEQ ID NOS: 1-8 and 18-47 described above that additionally consist of a protein, polypeptide, or polynucleotide encoding a sequence having at least about 70% or more identity and/or conservative substitutions to amino acid residues 302-315 of SEQ ID NO: 2 when such sequence is aligned with SEQ ID NO: 2. In another embodiment, the described phosphatases and homologues thereof consist of a protein, polypeptide, or polynucleotide encoding a protein having about 80% or more identity and/or conservative substitutions to amino acid residues 302-315 of SEQ ID NO: 2 when such sequence is aligned with SEQ ID NO: 2. In another embodiment, the described phosphatases and homologues consist of a protein, polypeptide, or polynucleotide encoding a sequence of amino acid residues having at least about 70% or more identity to the sequence HIGDI-SYARGYSW (SEQ ID NO: 56). In another embodiment, the described phosphatases and homologues thereof consist of a protein, polypeptide, or polynucleotide encoding a sequence of amino acid residues having the sequence HIGDISYAR-GYSW (SEQ ID NO: 56) or a protein, polypeptide, or polynucleotide encoding a protein having a homologous sequence to the foregoing sequences with only conservative substitutions, as described above, to those sequences.

In another embodiment, the described phosphatases and about 340, at least about 360, from about 250 to about 660, 50 homologues thereof consist of a protein, polypeptide, or polynucleotide encoding a sequence of amino acid residues having at least about 70% or more identity to the sequences KEKLTVSFVGNHDGEVHD (SEQ ID NO: 57), KERLTL-SYVGNHDGEVHD (SEQ ID NO: 58), REKLTLTYVGN-HDGQVHD (SEQ ID NO: 59), and KEKLTLTYIGNH-DGQVHD (SEQ ID NO: 60). In still yet another embodiment, the described phosphatases and homologues thereof consist of a protein, polypeptide, or polynucleotide encoding a sequence of amino acid residues having one or more of the sequences KEKLTVSFVGNHDGEVHD (SEQ ID NO: 57), KERLTLSYVGNHDGEVHD (SEQ ID NO 58), REKLTLTYVGNHDGQVHD (SEQ ID NO: 59), and KEKLTLTYIGNHDGQVHD (SEQ ID NO: 60) or a protein, polypeptide, or polynucleotide encoding a protein having a homologous sequence to one of the foregoing sequences with only conservative substitutions, as described above, to those sequences.

In a further embodiment, the described phosphatases and homologues thereof consist of a protein, polypeptide, or polynucleotide encoding a sequence of amino acid residues having the sequence (F/Y)(V/I)GNHDGXXH (SEQ ID NOS: 61-64), where the first residue of the sequence can be 5 F or Y and the second residue of the sequence can be V or I. In a still further embodiment, the described phosphatases and homologues thereof consist of a protein, polypeptide, or polynucleotide encoding a sequence of amino acid residues having the sequence (F/Y)(V/I)GNHDGXXH (SEQ ID NOS: 61-64), where the first residue of the sequence can be F or Y and the second residue of the sequence can be V or I, or a protein, polypeptide, or polynucleotide encoding a protein having a homologous sequence to the foregoing sequence with only conservative substitutions, as described 15 above, to the foregoing sequence. In a yet still further embodiment, the described phosphatases and homologues thereof consist of a protein, polypeptide, or polynucleotide encoding a sequence of amino acid residues having the sequence (F/Y)(V/I)GNHDGXXH (SEQ ID NOS: 61-64), 20 where the first residue of the sequence can be F or Y and the second residue of the sequence can be V or I, or a protein, polypeptide, or polynucleotide encoding a protein having a homologous sequence having at least about 70% identity and/or conservative substitution, as described above, to the 25 foregoing sequence.

In one embodiment, the described phosphatases and homologues thereof consist of a protein, polypeptide, or polynucleotide encoding a sequence of amino acid residues having at least about 60% or more identity and/or conser- 30 vative substitutions to amino acid residues 614-636 of SEQ ID NO: 2 (SEQ ID NO: 65) and/or having at least about 60% or more identity and/or conservative substitutions to the sequence of 23 amino acid residues of SEQ ID NOS: 4, 6, 8, 19, 21, 23, 25, 27, 29, 31, 33 and 47 aligned with residues 35 614-636 of SEQ ID NO: 2 (SEQ ID NO: 65), and where amino acid residues aligned with amino acid residues 614-636 of SEQ ID NO: 2 are predicted to form a transmembrane-like C-terminal motif by TMHMM analysis (http:// the described phosphatases and homologues thereof consist of a protein, polypeptide, or polynucleotide encoding a sequence of amino acid residues having at least about 70% or more identity and/or conservative substitutions to amino acid residues 614-636 of SEQ ID NO: 2 (SEQ ID NO: 65) 45 and/or having at least about 60% or more identity and/or conservative substitutions to the sequence of 23 amino acid residues of SEQ ID NOS: 4, 6, 8, 19, 21, 23, 25, 27, 29, 31, 33 and 47 aligned with residues 614-636 of SEQ ID NO: 2 (SEQ ID NO: 65), and where amino acid residues aligned 50 with amino acid residues 614-636 of SEQ ID NO: 2 (SEQ ID NO: 65) are predicted to form a transmembrane-like C-terminal motif by TMHMM analysis (http://www.cbs.dtu.dk/services/TMHMM/). In one embodiment, the described phosphatases and homologues thereof consist of a 55 protein, polypeptide, or polynucleotide encoding a sequence of amino acid residues having at least about 80% or more identity and/or conservative substitutions to amino acid residues 614-636 of SEQ ID NO: 2 and/or having at least about 60% or more identity and/or conservative substitu- 60 tions to the sequence of 23 amino acid residues of SEQ ID NOS: 4, 6, 8, 19, 21, 23, 25, 27, 29, 31, 33 and 47 aligned with residues 614-636 of SEQ ID NO: 2 (SEQ ID NO: 65), and where amino acid residues aligned with amino acid residues 614-636 of SEQ ID NO: 2 (SEQ ID NO: 65) are 65 predicted to form a transmembrane-like C-terminal motif by TMHMM analysis (http://www.cbs.dtu.dk/services/TM-

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HMM/). In one embodiment, the described phosphatases and homologues thereof consist of a protein, polypeptide, or polynucleotide encoding a sequence of amino acid residues having at least about 90% or more identity and/or conservative substitutions to amino acid residues 614-636 of SEQ ID NO: 2 and/or having at least about 90% or more identity and/or conservative substitutions to the sequence of 23 amino acid residues of SEQ ID NOS: 4, 6, 8, 19, 21, 23, 25, 27, 29, 31, 33 and 47 aligned with residues 614-636 of SEQ ID NO: 2, and where amino acid residues aligned with amino acid residues 614-636 of SEQ ID NO: 2 are predicted to form a transmembrane-like C-terminal motif by TMHMM analysis (http://www.cbs.dtu.dk/services/TM-HMM/).

In one embodiment, the described phosphatases or phosphatase genes consist of a protein, polypeptide, or polynucleotide encoding the sequence (L/M/V)-(L/MN)-Z-(G/ A)-(V/A/L)-Z-Z-G-(F/Y)-X-Z-G (SEQ ID NO: 66), where Z is any of the hydrophobic residues L, I, V, F, and M. In another embodiment, the described phosphatase or phosphatase genes consist of a protein, polypeptide, or polynucleotide encoding the sequence (L/M/V)-(L/MN)-Z-(G/ A)-(V/A/L)-Z-Z-G-(F/Y)-X-Z-G (SEQ ID NO: 66), or a protein, polypeptide, or polynucleotide encoding a sequence having at least 70% identity and/or conservative substitution to the foregoing sequence.

Embodiments also encompass derivatives of the disclosed polypeptides. For example, but not by way of limitation, derivatives may include peptides or proteins that have been modified, e.g., by glycosylation, acetylation, pegylation, phosphorylation, amidation, derivatization by known protecting/blocking groups, proteolytic cleavage, linkage to a cellular ligand or other protein, etc. Any of numerous chemical modifications may be carried out by known techniques, including, but not limited to, specific chemical cleavage, acetylation, formylation, etc. Additionally, the derivative may contain one or more non-classical amino

In another aspect, an isolated nucleic acid molecule www.cbs.dtu.dk/services/TMHMM/). In one embodiment, 40 encodes a variant of a polypeptide in which the amino acid sequences have been modified by genetic engineering so that biological activities of the polypeptides are either enhanced or reduced, or the local structures thereof are changed without significantly altering the biological activities. In one aspect, these variants can act as either agonists or as antagonists. An agonist can retain substantially the same or a portion of the biological activities of the polypeptides and an antagonist can inhibit one or more of the activities of the polypeptides. Such modifications include amino acid substitution, deletion, and/or insertion. Amino acid modifications can be made by any method known in the art and various methods are available to and routine for those skilled in the art.

For example, mutagenesis may be performed in accordance with any of the techniques known in the art including, but not limited to, synthesizing an oligonucleotide having one or more modifications within the sequence of a given polypeptide to be modified. Site-specific mutagenesis can be conducted using specific oligonucleotide sequences which encode the nucleotide sequence containing the desired mutations in addition to a sufficient number of adjacent nucleotides in the polypeptide. Such oligonucleotides can serve as primers which can form a stable duplex on both sides of the deletion junction being traversed. Typically, a primer of about 15 to about 75 nucleotides or more in length is preferred, with about 10 to about 25 or more residues on both sides of the junction of the sequence being altered. A

number of such primers introducing a variety of different mutations at one or more positions can be used to generate a library of mutants.

The technique of site-specific mutagenesis is well known in the art, as described in various publications (e.g., Kunkel 5 et al., Methods Enzymol., 154:367-82, 1987, which is hereby incorporated by reference in its entirety). In general, site-directed mutagenesis is performed by first obtaining a single-stranded vector or melting apart of two strands of a double stranded vector which includes within its sequence a 10 DNA sequence which encodes the desired peptide. An oligonucleotide primer bearing the desired mutated sequence is prepared, generally synthetically. This primer is then annealed with the single-stranded vector, and subjected to DNA polymerizing enzymes such as T7 DNA poly- 15 merase, in order to complete the synthesis of the mutationbearing strand. Thus, a heteroduplex is formed wherein one strand encodes the original non-mutated sequence and the second strand bears the desired mutation. This heteroduplex vector is then used to transform or transfect appropriate 20 cells, such as E. coli cells, and clones are selected which include recombinant vectors bearing the mutated sequence arrangement. As will be appreciated, the technique typically employs a phage vector which exists in both a single stranded and double stranded form. Typical vectors useful in 25 site-directed mutagenesis include vectors such as the M13 phage. These phages are readily commercially available and their use is generally well known to those skilled in the art. Double stranded plasmids are also routinely employed in site directed mutagenesis which eliminates the step of transferring the gene of interest from a plasmid to a phage.

Alternatively, the use of PCR with commercially available thermostable enzymes such as Taq DNA polymerase may be used to incorporate a mutagenic oligonucleotide primer into an amplified DNA fragment that can then be 35 cloned into an appropriate cloning or expression vector. See, e.g., Tomic et al., *Nucleic Acids Res.*, 18(6):1656, 1987, and Upender et al., *Biotechniques*, 18(1):29-30, 32, 1995, for PCR-mediated mutagenesis procedures, which are hereby incorporated in their entireties. PCR employing a thermo-40 stable ligase in addition to a thermostable polymerase may also be used to incorporate a phosphorylated mutagenic oligonucleotide into an amplified DNA fragment that may then be cloned into an appropriate cloning or expression vector (see e.g., Michael, *Biotechniques*, 16(3):410-2, 1994, 45 which is hereby incorporated by reference in its entirety).

Other methods known to those skilled in art of producing sequence variants of a given polypeptide or a fragment thereof can be used. For example, recombinant vectors encoding the amino acid sequence of the polypeptide or a 50 fragment thereof may be treated with mutagenic agents, such as hydroxylamine, to obtain sequence variants.

Optionally, the amino acid residues to be modified are surface exposed residues. Additionally, in making amino acid substitutions, preferably the amino acid residue to be 55 substituted is a conservative amino acid substitution, for example, a polar residue is substituted with a polar residue, a hydrophilic residue with a hydrophilic residue, hydrophobic residue with a hydrophobic residue, a positively charged residue with a positively charged residue with a negatively charged residue. Moreover, the amino acid residue that can be modified is not highly or completely conserved across strains or species and/or is critical to maintain the biological activities of the protein.

Accordingly, included in the scope of the disclosure are nucleic acid molecules encoding a polypeptide of the inven18

tion that contains amino acid modifications that are not critical to its biological activity.

#### 5.3 Fusion Proteins

The present disclosure further encompasses fusion proteins in which the polypeptides or fragments thereof, are recombinantly fused or chemically conjugated (e.g., covalent and non-covalent conjugations) to heterologous polypeptides (i.e., an unrelated polypeptide or portion thereof, preferably at least 10, at least 20, at least 30, at least 40, at least 50, at least 60, at least 70, at least 80, at least 90 or at least 100 amino acids of the polypeptide) to generate fusion proteins. The fusion can be direct, but may occur through linker sequences.

In one aspect, the fusion protein comprises a polypeptide which is fused to a heterologous signal sequence at its N-terminus. For example, the signal sequence naturally found in the polypeptide can be replaced by a signal sequence which is derived from a heterologous origin. Various signal sequences are commercially available.

In another embodiment, a polypeptide can be fused to tag sequences, e.g., a hexa-histidine peptide, among others, many of which are commercially available. As described in Gentz et al., 1989, *Proc. Natl. Acad. Sci. USA*, 86:821-824, for instance, hexa-histidine provides for convenient purification of the fusion protein. Other examples of peptide tags are the hemagglutinin "HA" tag, which corresponds to an epitope derived from the influenza hemagglutinin protein (Wilson et al., 1984, *Cell*, 37:767) and the "flag" tag (Knappik et al., 1994, *Biotechniques*, 17(4):754-761). These tags are especially useful for purification of recombinantly produced polypeptides.

Fusion proteins can be produced by standard recombinant DNA techniques or by protein synthetic techniques, e.g., by use of a DNA synthesizer. For example, a nucleic acid molecule encoding a fusion protein can be synthesized by conventional techniques including automated DNA synthesizers. Alternatively, PCR amplification of gene fragments can be carried out using anchor primers which give rise to complementary overhangs between two consecutive gene fragments which can subsequently be annealed and reamplified to generate a chimeric gene sequence (see, e.g., Current Protocols in Molecular Biology, Ausubel et al., eds., John Wiley & Sons, 1992).

The nucleotide sequence coding for a fusion protein can be inserted into an appropriate expression vector, i.e., a vector which contains the necessary elements for the transcription and translation of the inserted protein-coding sequence.

In a specific embodiment, the expression of a fusion protein is regulated by an inducible promoter.

#### 5.4 Preparation of Transgenic Plants

Carbon flow is a key process in plant biology and high energy carbon molecules (e.g. glucose) were harvested by plant through photosynthesis. The carbon molecules were then converted into more complicated carbohydrate molecules such as starch, cellulose, etc. Cellulose is the major component of cell wall and starch is the major storage form of glucose in plant cells and plant seeds. Therefore, the efficiency and/or the equilibrium of the carbon flow process become a limiting factor for plant growth and crop yield.

The present disclosure is based upon the discovery that overexpression of a membrane-bound phosphatase can enhance the growth performance of plants by altering its carbon metabolism, as indicated by, for example, a faster growth rate, a higher sugar contents, and a higher seed yield.

In an embodiment, the present disclosure provides a transgenic plant containing a nucleic acid molecule that

encodes and expresses a phosphatase having a C-terminal transmembrane-like domain. The transgenic plants disclosed herein have faster growth rate, and higher seed yield to comparable unengineered plants i.e. same species (strain). In a specific embodiment, such a phosphatase is from a plant 5 species having a phosphatase activity and a C-terminal motif. In another embodiment, a transgenic plant disclosed herein comprises a nucleic acid molecule encoding phosphatase and expresses AtPAP2 (SEQ ID NO: 2) and/or other PAPs with a C-terminal motif including one or more of SEQ 10 ID NOS: 4, 6, 8, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45 and 47. In another embodiment, the phosphatase is expressed on cellular membrane, for example, the ER or the Golgi apparatus. Such a membrane expression of a phosphatase in plants can be achieved by fusing onto the 15 C-terminus with a nucleotide sequence encoding a C-terminal motif peptide which can efficiently attach the phosphatase upon translation thereof from the cells of a given plant. Accordingly, in another embodiment, a transgenic plant comprises a nucleic acid molecule encoding phos- 20 phatase and expresses AtPAP2 (SEQ ID NO: 2) and/or other PAPs with a C-terminal motif including SEQ ID NOS: 4, 6, 8, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45 and 47, except that all or a portion, particularly an N-terminal portion, of amino acid residues 1 to 80, preferably all or a 25 portion of amino acid residues 1 to 30, of SEQ ID NO: 2 or all or a portion, particularly an N-terminal portion, of amino acid residues 1 to 80, preferably all or a portion of amino acid residues 1 to 30, of SEQ ID NOS: 4, 6, 8, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45 or 47, are replaced 30 by a heterologous plant signal peptide by genetic engineering. In such a transgenic plant, the phosphatases are directed to various organelles/compartments of the cells. In another embodiment, a transgenic plant comprises a nucleic acid molecule encoding phosphatase and expresses homologues, 35 derivatives, and/or fragments thereof having at least one functional feature and/or structural feature of a phosphatase polypeptide. In all embodiments where all or a portion of the N-terminal portion of SEQ ID NOS: 4, 6, 8, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, 45 and/or 47 are replaced, 40 the embodiments include homologues to such sequences, as described above, having at least one functional feature and/or structural feature of a phosphatase polypeptide. In yet another embodiment, a transgenic plant comprises a nucleic acid molecule that hybridizes under stringent conditions, as 45 defined herein, to a nucleic acid molecule having the sequence of SEQ ID NOS: 1, 3, 5, 7, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44 or 46, or a complement thereof, and encodes a protein or polypeptide that exhibits at least one structural and/or functional feature of the disclosed 50 phosphatase polypeptides. Specifically, the production of transgenic plant that overexpressed a membrane-bound phosphatase, which contributes to improving plant physiology, such as plant growth rate and characteristics, for example, in seed yield, is provided.

Accordingly, also provided are chimeric gene constructs for genetic modification of plants to increase their growth rate and improve the yield. The chimeric gene constructs comprise a sequence that encodes substantially solely for a phosphatase enzyme that carry a C-terminal transmembrane-like motif. Such a phosphatase enzyme can be derived from the purple acid phosphatase family. In a specific embodiment, the chimeric gene constructs comprise a nucleic acid having the sequence of SEQ ID NOS: 1, 3, 5, 7, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44 or 46. 65 In another embodiment, the chimeric gene constructs comprise a nucleic acid molecule that encodes a homologue or

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fragment thereof having at least one functional feature and/or structural feature of a phosphatase polypeptide. In another specific embodiment, the chimeric gene constructs comprise a sequence that hybridizes under stringent conditions, as defined herein, to a nucleic acid having the sequence of SEQ ID NOS: 1, 3, 5, 7, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44 or 46, or a complement thereof, wherein the sequence encodes a protein or a polypeptide that exhibits at least one structural and/or functional feature of the phosphatase polypeptides. Furthermore, the phosphatases encoded by the nucleic acid molecules contained in the chimeric gene constructs can be any other phosphatases that have similar structural characteristics, such as having a C-terminal transmembrane-like motif, to those of the phosphatases described herein. Such phosphatase include, but not limited to, the following polypeptides: Purple acid phosphatases from Zea mays (Accession No: ACG47621); and Oryza sativa (Accession No: BAC15853.1).

The phosphatase-coding sequence is operatively linked to upstream and downstream regulatory components, preferably heterologous to the phosphatase sequence; for example CMV 35S promoter, which acts to cause expression of the gene (production of the enzyme) in plant cells (see Section 6.2). When a construct containing a gene for a phosphatase according to this disclosure, is introduced into plant cells by a conventional transformation method, such as microparticle bombardment, Agrobacterium infection, or microinjection, the gene is expressed in the cells under the control of the regulatory sequences. The expressed phosphatase successfully interacts with the biosynthetic machinery that is naturally present in the plant cells to alter the carbon metabolism. By altering the carbon metabolism, the method described herein also favors the growth rate of the plant, resulting in faster growth rate and higher yield. Thus, the time required for the maturation of the plant and the time required for flowering is shortened. Also provided are methods for increasing growth rate and yield of plants, comprising the step of inserting into such plant cells or the cells of such whole plants a chimeric gene construct.

In specific embodiments, *Arabidopsis* (see Section 6) was adopted as the model system. An overexpression construct the gene coding for phosphatase were introduced into *Arabidopsis*.

In an embodiment, the phosphatase from *Arabidopsis* is used. The results obtained with this disclosure indicate that the growth rate and the seed yield of transgenic *Arabidopsis* were enhanced by overexpressing this gene (see Section 6.5 and FIG. **8** and Table 3).

While any plant species can be modified using the expression cassette and methods described herein, preferably included without limitation are species from the following genera with representative species in parentheses:

Monocots: genera Asparagus (asparagus), Bromus (cheatgrass), Hemerocallis (daylily), Hordeum (barley), 55 Lolium (ryegrass), Oryza (rice), Panicum (Switchgrass), Pennisetum (fountaingrass), Saccharum (Sugar cane), Sorghum, Trigonella (fenu grass), Triticum (wheat), Zea (corn); and

Dicots: genera Antirrhinum (flower sp.), Arabidopsis (thaliana), Arachis (peanut), Atropa (deadly nightshade), Brassica (rapeseed), Browallia, Capsicum (pepper), Carthamus (safflower), Cichorium (chicory), Citrus (orange, lemon), Chrysanthemum, Cucumis (cucumber), Datura (thorn apple), Daucus (carrot), Digitalis (foxglove), Fragaria (strawberry), Geranium (flower sp.), Glycine (soybean), Helianthus (sunflower), Hyscyamus, Ipomoea (morning glory), Latuca (lettuce), Linum (linseed), Lotus (flower

sp.), Lycopersicon (tomato), Majorana, Malva (cotton), Manihot, Medicago (alfalfa), Nemesia, Nicotiana (tobacco), Onobrychis, Pelargonium (citrosa), Petunia (flower sp.), Ranunculus (flower sp.), Raphanus (radishes), Salpiglossis, Senecio (flower sp.), Sinapis (albae semen), Solanum (potato), Trifolium (clovers), Vigna (mungbean, faba bean), Vitis (grape).

Genetic engineering of plants can be achieved in several ways. The most common method is Agrobacterium-mediated transformation. In this method, A. tumefaciens, which in nature infects plants by inserting tumor causing genes into a plant's genome, is altered. Selected genes are engineered into the T-DNA of the bacterial Ti (tumor-inducing) plasmid of A. tumefaciens in laboratory conditions so that they 15 become integrated into the plant chromosomes when the T-DNA is transferred to the plant by the bacteria's own internal transfer mechanisms. The only essential parts of the T-DNA are its two small (25 base pair) border repeats, at least one of which is needed for plant transformation. The 20 bacterial genes encoding for plant hormones that promote tumor growth are excised from the T-DNA and replaced with a sequence of DNA that typically contains: a selectable marker (e.g. an antibiotic-resistance gene; usually kanamycin resistance), a restriction site—a site with a specific 25 sequence of nucleotides where a restriction enzyme will cut the DNA, and the desired genes to be incorporated into the plant (B. Tinland, 1996. The integration of T-DNA into plant genomes. Trends in Plant Science 1, 178-184; D. Grierson (ed.) 1991. Plant Genetic Engineering. Blackie, Glasgow). 30 Agrobacterium can be added to plant protoplasts (plant cells with cell walls removed) in culture, that are then allowed to regenerate cell walls at which point non-transformed plants are killed with antibiotics for which the transformed plants have been given resistance genes. Plantlets are then regen- 35 erated from the surviving transformed cells using standard plant tissue culture techniques. In an alternative technique, sterile disks or fragments of vegetative portions of plants are place in liquid culture medium with Agrobacterium, then hormones are used to induce rooting thereby regenerate 40 plantlets which are grown on selection media. A third technique for delivering genes is possible for some plants such as Arabidopsis where the Agrobacterium or even "naked" DNA can be infused through the seed coat to cause transformation (Clough S J and Bent A F, 1998. Floral dip: 45 a simplified method for Agrobacterium-mediated transformation of Arabidopsis thaliana. Plant J 16:735-43).

The biolistic method for genetic engineering of plants was developed more recently and is becoming more widely employed. In this method, very small particles (micropro- 50 jectiles) of tungsten or gold coated with biologically active DNA are propelled at high-velocities into plant cells using an electrostatic pulse, air pressure, or gunpowder percussion. As the particles pass through the cell, the DNA dissolves and can then integrate into the genome of that cell 55 and its progeny. It has been demonstrated this method can produce stable transformants (Christou, P., et al., 1988. Stable transformation of soybean callus by DNA-coated gold particles, *Plant Physiology* 87:671-674). The method can be practiced on whole plants and is particularly effective 60 on meristematic tissue. It is also capable of delivering DNA either to the nucleus or into mitochondria (Johnston, S. A., et al., 1988. Mitochondrial transformation in yeast by bombardment with microprojectiles (Science 240, 1538-41) and chloroplasts (Svab, Z., et al., 1990, Stable transformation of 65 plastids in higher plants, Proc Natl Acad. Sci. USA 87, 8526-8530).

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The electroporation method of plant genetic engineering has met with less success. In this technique, protoplasts in culture take up pure DNA when treated with certain membrane-active agents or with electroporation, a rapid pulse of high-voltage direct current. Once the DNA has entered the protoplast it can be integrated into the cells genome. Standard tissue culture techniques are then used to regenerate transgenic plants.

The microinjection method of plant genetic engineering is perhaps the most difficult. In this method, DNA is microinjected into target plant cells using very thin glass needles in a method similar to that used with animals. The technique is laborious, ineffective, and impractical for generating large numbers of transgenic plants.

The method chosen for genetically engineering plants is most often dependent on the targeted plant species and which methods have been proven effective therein.

#### 5.5 Preparation of Antibodies

Antibodies which specifically recognize one of the described phosphatase polypeptides or fragments thereof can be used for detecting, screening, and isolating the polypeptide of the invention or fragments thereof, or similar sequences that might encode similar enzymes from the other organisms. For example, in one specific embodiment, an antibody which immunospecifically binds AtPAP2 or fragments thereof can be used for various in vitro detection assays, including enzyme-linked immunosorbent assays (ELISA), radioimmunoassays, Western blot, etc., for the detection of the polypeptide of the invention or fragments, derivatives, homologues, or variants thereof, or similar molecules having the similar enzymatic activities as the phosphatase polypeptides, in samples, for example, a biological material, including plant cells, plants, food, drinks, or any materials derived from plants.

Antibodies specific for the described phosphatase polypeptides can be generated by any suitable method known in the art. Polyclonal antibodies to an antigen-of-interest can be produced by various procedures well known in the art. For example, an antigen derived from the phosphatase polypeptide can be administered to various host animals including, but not limited to, rabbits, mice, rats, etc., to induce the production of antisera containing polyclonal antibodies specific for the antigen. Various adjuvants may be used to increase the immunological response, depending on the host species, and include but are not limited to, Freund's (complete and incomplete) adjuvant, mineral gels such as aluminum hydroxide, surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, keyhole limpet hemocyanins, dinitrophenol, and potentially useful adjuvants for humans such as BCG (Bacille Calmette-Guerin) and Corynebacterium parvum. Such adjuvants are also well known in the art.

Monoclonal antibodies can be prepared using a wide variety of techniques known in the art including the use of hybridoma, recombinant, and phage display technologies, or a combination thereof. For example, monoclonal antibodies can be produced using hybridoma techniques including those known in the art and taught, for example, in Harlow et al., *Antibodies: A Laboratory Manual*, (Cold Spring Harbor Laboratory Press, 2nd ed. 1988); Hammerling, et al., in: *Monoclonal Antibodies and T-Cell Hybridomas*, pp. 563-681 (Elsevier, N.Y., 1981) (both of which are incorporated by reference in their entireties). The term "monoclonal antibody" as used herein is not limited to antibodies produced through hybridoma technology. The term "monoclonal antibody" refers to an antibody that is derived from a

single clone, including any eukaryotic, prokaryotic, or phage clone, and not the method by which it is produced.

Methods for producing and screening for specific antibodies using hybridoma technology are routine and well known in the art. In a non-limiting example, mice can be 5 immunized with an antigen of interest or a cell expressing such an antigen. Once an immune response is detected, e.g., antibodies specific for the antigen are detected in the mouse serum, the mouse spleen is harvested and splenocytes isolated. The splenocytes are then fused by well known techniques to any suitable myeloma cells. Hybridomas are selected and cloned by limiting dilution. The hybridoma clones are then assayed by methods known in the art for cells that secrete antibodies capable of binding the antigen. Ascites fluid, which generally contains high levels of antibodies, can be generated by inoculating mice intraperitoneally with positive hybridoma clones.

Antibody fragments which recognize specific epitopes may be generated by known techniques. For example, Fab and F(ab')<sub>2</sub> fragments may be produced by proteolytic cleavage of immunoglobulin molecules, using enzymes such as papain (to produce Fab fragments) or pepsin (to produce F(ab')<sub>2</sub> fragments). F(ab')<sub>2</sub> fragments contain the complete light chain, and the variable region, the CH1 region and the hinge region of the heavy chain.

The antibodies or fragments thereof can be also produced by any method known in the art for the synthesis of antibodies, in particular, by chemical synthesis or preferably, by recombinant expression techniques.

The nucleotide sequence encoding an antibody may be 30 obtained from any information available to those skilled in the art (i.e., from Genbank, the literature, or by routine cloning). If a clone containing a nucleic acid encoding a particular antibody or an epitope-binding fragment thereof is not available, but the sequence of the antibody molecule or 35 epitope-binding fragment thereof is known, a nucleic acid encoding the immunoglobulin may be chemically synthesized or obtained from a suitable source (e.g., an antibody cDNA library, or a cDNA library generated from, or nucleic acid, preferably poly A+ RNA, isolated from any tissue or 40 cells expressing the antibody, such as hybridoma cells selected to express an antibody) by PCR amplification using synthetic primers hybridizable to the 3' and 5' ends of the sequence or by cloning using an oligonucleotide probe specific for the particular gene sequence to identify, e.g., a 45 cDNA clone from a cDNA library that encodes the antibody. Amplified nucleic acids generated by PCR may then be cloned into replicable cloning vectors using any method well known in the art.

Once the nucleotide sequence of the antibody is determined, the nucleotide sequence of the antibody may be manipulated using methods well known in the art for the manipulation of nucleotide sequences, e.g., recombinant DNA techniques, site directed mutagenesis, PCR, etc. (see, for example, the techniques described in Sambrook et al., 55 supra; and Ausubel et al., eds., 1998, *Current Protocols in Molecular Biology*, John Wiley & Sons, NY, which are both incorporated by reference herein in their entireties), to generate antibodies having a different amino acid sequence by, for example, introducing amino acid substitutions, deletions, and/or insertions into the epitope-binding domain regions of the antibodies or any portion of antibodies which may enhance or reduce biological activities of the antibodies.

Recombinant expression of an antibody requires con- 65 struction of an expression vector containing a nucleotide sequence that encodes the antibody. Once a nucleotide

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sequence encoding an antibody molecule or a heavy or light chain of an antibody, or portion thereof has been obtained, the vector for the production of the antibody molecule may be produced by recombinant DNA technology using techniques well known in the art as discussed in the previous sections. Methods which are well known to those skilled in the art can be used to construct expression vectors containing antibody coding sequences and appropriate transcriptional and translational control signals. These methods include, for example, in vitro recombinant DNA techniques, synthetic techniques, and in vivo genetic recombination. The nucleotide sequence encoding the heavy-chain variable region, light-chain variable region, both the heavy-chain and light-chain variable regions, an epitope-binding fragment of the heavy- and/or light-chain variable region, or one or more complementarity determining regions (CDRs) of an antibody may be cloned into such a vector for expression. Thus-prepared expression vector can be then introduced into appropriate host cells for the expression of the antibody. Accordingly, embodiments include host cells containing a polynucleotide encoding an antibody specific for the disclosed phosphatase polypeptides or fragments thereof.

The host cell can be co-transfected with two expression vectors, the first vector encoding a heavy chain derived polypeptide and the second vector encoding a light chain derived polypeptide. The two vectors may contain identical selectable markers which enable equal expression of heavy and light chain polypeptides or different selectable markers to ensure maintenance of both plasmids. Alternatively, a single vector may be used which encodes, and is capable of expressing, both heavy and light chain polypeptides. In such situations, the light chain should be placed before the heavy chain to avoid an excess of toxic free heavy chain (Proudfoot, 1986, *Nature*, 322:52; and Kohler, 1980, *Proc. Natl. Acad. Sci. USA*, 77:2197). The coding sequences for the heavy and light chains may comprise cDNA or genomic DNA.

In another embodiment, antibodies can also be generated using various phage display methods known in the art. In phage display methods, functional antibody domains are displayed on the surface of phage particles which carry the polynucleotide sequences encoding them. In a particular embodiment, such phage can be utilized to display antigen binding domains, such as Fab and Fv or disulfide-bond stabilized Fvs, expressed from a repertoire or combinatorial antibody library (e.g., human or murine). Phage expressing an antigen binding domain that binds the antigen of interest can be selected or identified with antigen, e.g., using labeled antigen or antigen bound or captured to a solid surface or bead. Phages used in these methods are typically filamentous phage, including fd and M13. The antigen binding domains are expressed as a recombinantly fused protein to either the phage gene III or gene VIII protein. Examples of phage display methods that can be used to make the immunoglobulins, or fragments thereof, include those disclosed in Brinkman et al., 1995, J. Immunol. Methods 182:41-50; Ames et al., 1995, J. Immunol. Methods 184:177-186; Kettleborough et al., 1994, Eur. J. Immunol., 24:952-958; Persic et al., 1997, Gene, 187:9-18; Burton et al., 1994, Advances in Immunology 57:191-280; PCT application No. PCT/GB91/01134; PCT publications WO 90/02809; WO 91/10737; WO 92/01047; WO 92/18619; WO 93/11236; WO 95/15982; WO 95/20401; and U.S. Pat. Nos. 5,698,426; 5,223,409; 5,403,484; 5,580,717; 5,427,908; 5,750,753; 5,821,047; 5,571,698; 5,427,908; 5,516,637; 5,780,225; 5,658,727; 5,733,743 and 5,969,108; each of which is incorporated herein by reference in its entirety.

As described in the above documents, after phage selection, the antibody coding regions from the phage can be isolated and used to generate whole antibodies, including human antibodies, or any other desired fragments, and expressed in any desired host, including mammalian cells, 5 insect cells, plant cells, yeast, and bacteria, e.g., as described in detail below. For example, techniques to recombinantly produce Fab, Fab' and F(ab)2 fragments can also be employed using methods known in the art such as those disclosed in PCT publication WO 92/22324; Mullinax et al., 10 1992, Bio Techniques 12(6):864-869; and Sawai et al., 1995, AJRI 34:26-34; and Better et al., Science, 240:1041-1043, 1988 (each of which is incorporated by reference in its entirety). Examples of techniques which can be used to produce single-chain Fvs and antibodies include those 15 described in U.S. Pat. Nos. 4,946,778 and 5,258,498; Huston et al., 1991, Methods in Enzymology 203:46-88; Shu et al., 1993, PNAS 90:7995-7999; and Skerra et al., 1988, Science 240:1038-1040.

Once an antibody molecule has been produced by any 20 methods described above, it may then be purified by any method known in the art for purification of an immunoglobulin molecule, for example, by chromatography (e.g., ion exchange, affinity, particularly by affinity for the specific antigen after Protein A or Protein G purification, and sizing 25 column chromatography), centrifugation, differential solubility, or by any other standard techniques for the purification of proteins. Further, the antibodies or fragments thereof may be fused to heterologous polypeptide sequences described herein or otherwise known in the art to facilitate 30 purification.

Antibodies fused or conjugated to heterologous polypeptides may be used in in vitro immunoassays and in purification methods (e.g., affinity chromatography) well known in the art. See e.g., PCT publication Number WO 93/21232; 35 EP 439,095; Naramura et al., 1994, *Immunol. Lett.* 39:91-99; U.S. Pat. No. 5,474,981; Gillies et al., 1992, PNAS 89:1428-1432; and Fell et al., 1991, *J. Immunol.* 146:2446-2452, which are incorporated herein by reference in their entireties.

Antibodies may also be attached to solid supports, which are particularly useful for immunoassays or purification of the described polypeptides or fragments, derivatives, homologues, or variants thereof, or similar molecules having the similar enzymatic activities as the polypeptide of the invention. Such solid supports include, but are not limited to, glass, cellulose, polyacrylamide, nylon, polystyrene, polyvinyl chloride or polypropylene.

#### 5.6 Detection Assays

An exemplary method for detecting the presence or 50 absence of an overexpressed phosphatase polypeptide or an inserted phosphatase-encoding nucleic acid in a biological sample involves obtaining a biological sample from various sources and contacting the sample with a compound or an agent capable of detecting a polypeptide or nucleic acid 55 (e.g., mRNA, genomic DNA) such that the presence of a heterologous polypeptide or nucleic acid is detected in the sample. An exemplary agent for detecting mRNA or genomic DNA encoding an inserted phosphatase polypeptide is a labeled nucleic acid probe capable of hybridizing to 60 mRNA or genomic DNA encoding any of the described phosphatase polypeptides. The nucleic acid probe can be, for example, a full-length cDNA, such as the nucleic acid of SEQ ID NOS: 1, 3, 5, 7, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44 or 46, or a portion thereof, such as an 65 oligonucleotide of at least one of at least about 15, at least about 20, at least about 25, at least about 30, at least about

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50, at least about 100, at least about 250, at least about 500, or more nucleotides in length and sufficient to specifically hybridize under stringent conditions to a mRNA or genomic DNA encoding a polypeptide of the invention.

An exemplary agent for detecting an over-expressed phosphatase polypeptide is an antibody capable of binding to a phosphatase polypeptide product of an inserted phosphatase gene, preferably an antibody with a detectable label. Antibodies can be polyclonal, or more preferably, monoclonal. An intact antibody, or a fragment thereof (e.g., Fab or  $F(ab')_2$ ) can be used. See also the detailed descriptions about antibodies in Section 5.5.

The term "labeled", with regard to the probe or antibody, is intended to encompass direct labeling of the probe or antibody by coupling (i.e., physically linking) a detectable substance to the probe or antibody, as well as indirect labeling of the probe or antibody by reactivity with another reagent that is directly labeled. Examples of indirect labeling include detection of a primary antibody using a fluorescently labeled secondary antibody and end-labeling of a DNA probe with biotin such that it can be detected with fluorescently labeled streptavidin. The detection method can be used to detect mRNA, protein, or genomic DNA in a sample in vitro as well as in vivo. For example, in vitro techniques for detection of mRNA include Northern hybridizations and in situ hybridizations. In vitro techniques for detection of a heterologous polypeptide include enzyme linked immunosorbent assays (ELISAs), Western blots, immunoprecipitations and immunofluorescence. In vitro techniques for detection of genomic DNA include Southern hybridizations. Furthermore, in vivo techniques for detection of a heterologous polypeptide include introducing into a subject organism a labeled antibody directed against the polypeptide. For example, the antibody can be labeled with a radioactive marker whose presence and location in the subject organism can be detected by standard imaging techniques, including autoradiography.

In a specific embodiment, the methods further involve obtaining a control sample from a control subject, contacting the control sample with a compound or agent capable of detecting an over-expressed polypeptide product or the mRNA transcription product or genomic DNA encoding an inserted phospatase gene, such that the presence of the polypeptide or mRNA or genomic DNA encoding the phosphatase polypeptide is detected in the sample, and comparing the presence of the phosphatase polypeptide or mRNA or genomic DNA encoding the polypeptide in the control sample with the presence of the polypeptide or mRNA or genomic DNA encoding endogenous phosphatase polypeptides in the test sample.

Embodiments also encompass kits for detecting the presence of a heterologous polypeptide or nucleic acid in a test sample.

The kit, for example, can comprise a labeled compound or agent capable of detecting the polypeptide or mRNA encoding the polypeptide in a test sample and means for determining the amount of the polypeptide or mRNA in the sample (e.g., an antibody which binds the polypeptide or an oligonucleotide probe which binds to DNA or mRNA encoding the polypeptide). Kits can also optionally include instructions for use.

For antibody-based kits, the kit can comprise, for example: (1) a first antibody (e.g., attached to a solid support) which binds to a phosphatase polypeptide; and, optionally, (2) a second, different antibody which binds to either the polypeptide or the first antibody and is conjugated to a detectable agent.

For oligonucleotide-based kits, the kit can comprise, for example: (1) an oligonucleotide, e.g., a detectably labeled oligonucleotide, which hybridizes to a nucleic acid sequence encoding an inserted phosphatase polypeptide or (2) a pair of primers useful for amplifying a nucleic acid molecule encoding an inserted phosphatase polypeptide. The kit can also comprise, e.g., a buffering agent, a preservative, or a protein stabilizing agent. The kit can also comprise components necessary for detecting the detectable agent (e.g., an enzyme or a substrate). The kit can also contain a control sample or a series of control samples which can be assayed and compared to the test sample contained. Each component of the kit is usually enclosed within an individual container and all of the various containers are within a single package along with instructions for use.

#### 5.7 Commercial Application of Transgenic Plants

The transgenic plants generated can have many useful applications, including food, feed, biomass, biofuels (starch, cellulose, seed lipids) and wood pulp. The enhanced growth rate of the transgenic plants may provide additional carbon <sup>20</sup> dioxide fixation per hectare of land per year and thus generate carbon credits.

#### 6. EXAMPLES

The following examples illustrate the cloning of AtPAP2, its overexpression in transgenic *Arabidopsis*, and the characterization of the transgenic plants. These examples should not be construed as limiting. The following examples illustrate some embodiments. Unless otherwise indicated in the following examples and elsewhere in the specification and claims, all parts and percentages are by weight, all temperatures are in degrees Centigrade, and pressure is at or near atmospheric pressure.

#### 6.1 Sequence Alignment and Phylogenetic Analysis

PAP2 locus and its genomic organization, including its intron/exon boundaries, were identified in the *Arabidopsis* Col-0 ecotype (http://www.arabidopsis.org). Sequence 40 alignment and phylogenetic tree were conducted using MEGA4 (Kumar et al., 2004) and ClustalW program (http://www.ebi.ac.uk/Tools/clustalw2/index.html). Amino acid sequence comparisons were performed using CLC Sequence Viewer 5.1.1 (www.cicbio.com).

Twenty nine PAP-like sequences were identified from the *Arabidopsis* genome and a phylogenetic tree was produced by neighbor-joining algorithm (FIG. 1). The gene locus of AtPAP2 (At1g13900) composes of two exons and the coding region is 1971 bp in length (SEQ ID NO: 1), which is 50 predicted to encode a polypeptide of ~73.7-KD. Among the twenty nine PAP-like protein sequences, only AtPAP2 and AtPAP9 carry a unique hydrophobic motif at their C-termini by TMHMM analysis (http://www.cbs.dtu.dk/services/TM-HMM-2.0/) (FIG. 3). AtPAP2 was found to share 72% 55 sequence identity in amino acid sequence with AtPAP9. Two sequences from *Zea mays* (Accession No: ACG47621) and *Oryza sativa* (Accession No: BAC15853.1) were found to share 58% and 57% a.a. identity with AtPAP2, respectively. Their sequences were aligned in FIG. 2.

AtPAP2-like sequences from other plant species that carry a hydrophobic motif at their C-termini were retrieved by tblastn program from Plant GDB database (http://www.plantgdb.org/) and NCBI database (http://blast.ncbi.n-lm.nih.gov/Blast.cgi) using the amino acid sequence of 65 AtPAP2 as the search sequence. cDNA and protein sequences that share high homology with that of AtPAP2

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were identified in Zea mays (SEQ ID NOs: 7 and 8), Brassica rapa (SEQ ID NOs: 18 and 19), Hordeum vulgare (SEQ ID NOs: 20 and 21), Medicago truncatula (SEQ ID NOs: 22 and 23), Physcomitrella patens (SEQ ID NOs: 24 and 25), Populus trichocarpa (SEQ ID NOs: 26 and 27), Saccharum officinarum (SEQ ID NOs: 28 and 29), Solanum tuberosum (SEQ ID NOs: 30 and 31), Vitis vinifera (SEQ ID NOs: 32 and 33), Oryza sativa (SEQ ID NOs: 34 and 35), Gossypium hirsutum (SEQ ID NOs: 36 and 37) Panicum virgatum (SEQ ID NOs: 38 and 39), Solanum lycopersicum (SEQ ID NOs: 40 and 41), Sorghum bicolor (SEQ ID NOs: 42 and 43) and Triticum aestivum (SEQ ID NOs: 44 and 45).

The cDNA sequences of AtPAP-like sequences were amplified from a local *Glycine max* variety (SEQ ID NO: 5) and the *Brassica napus* cultivar Westar (SEQ ID NO: 46) by RT-PCR using primers designed from corresponding EST sequences, which were retrieved from the Plant GDB database (http://www.plantgdb.org/).

#### 6.2 Screening of T-DNA Line and Production of Overexpression Lines and Complementation Lines in *Arabidopsis*

T-DNA insertion lines of PAP2 gene (Arabidopsis 25 genomic locus name: Salk 013567), in the Col ecotype were obtained from Arabidopsis Biological Resources Center (Alonso et al., 2003). Homologous T-DNA lines were identified by genomic PCR screening from SIGnAl database (http://signal.salk.edu/cgi-bin/tdnaexpress) by using the primers (LBa1, 5'-TGGTTCACGTAGTGGGCCATCG-3', SEQ ID NO: 9) and PAP2 specific forward primer (P2LP, 5'-TTGAAGTTTAACATGCCTGGG-3, SEQ ID NO: 10) and reverse primer (P2RP, 5'-TCCAATGCTCGA TTGATT-AGC-3', SEQ ID NO: 11). The PCR product was sequenced 35 and the T-DNA insertion site was confirmed. To exclude the possibility that another T-DNA locus interferes with the PAP2 mutant site, homologous pap2 mutant lines were backcrossed to the wild-type to dilute the potential T-DNA sites. The produced heterozygous pap2 mutants were grown on the MS plates containing 50 mg/ml Kanamycin. The ratio of the resistant to sensitive plants was about 3:1. These results demonstrated a single insertion locus site of the T-DNA line (pap2-8) lines.

The inability of the T-DNA line to express full length AtPAP2 mRNA was confirmed by RT-PCR. Total RNA was extracted from 10-day-old seedlings grown on MS with 2% (w/v) sucrose using the ThIzol RNA isolation method (Invitrogen) with DNase I treatment. cDNAs were generated using Superscript III reverse transcriptase (Invitrogen, Carlsbad, Calif., USA) using an oligo dT primer. Two gene-specific primers, P2YF (5'-GGCCGTCGACAT-GATCGTTAATT TCTCTTTC-3'SEQ ID NO: 12) and (5'-CCGGACTAGTTCATGTCTCCTCGTTCTT-GAC-3' SEQ ID NO: 13), were used to amplify a 1971 bp coding region of AtPAP2. For each sample, 1 µg of cDNA was amplified for 30 cycles, with an annealing temperature of 50° C. and using elongation factor (EF) primers, EF-1 (5'-GTTTCACATCAACATTGTGGTCA TTGG-3, SEQ ID NO: 14) and EF-2 (5'-GAGTACTTGGGGGTAGTG-60 GCATCC-3, SEQ ID NO: 15) (Axelos et al., 1989) for control experiment.

The inability of the T-DNA line to express protein was confirmed by Western blotting analysis (FIG. 4). Antiserum specific to AtPAP2 was raised in rabbit as described in Section 6.3.

To create transgenic AtPAP2 overexpressing lines or expressing this gene in the knockout mutants, the full length

coding region of the AtPAP2 cDNA was amplified by PCR using primers P2YF (SEQ ID NO: 12) and P2NR (SEQ ID NO: 13). A SalI site and a SpeI site were engineered into P2YF and P2NR, respectively. The resulting product (1976 bp) was inserted into the XhoI/Spe I sites of a binary vector, immediately downstream to the cauliflower mosaic virus (CaMV) 35S promoter (FIG. 5).

The vector was introduced into *Agrobacterium tumefaciens* strain GV3101 and then transformed by the floral dip method (Clough and Bent, 1998), into wild-type Col-0 to generate PAP2-overexpressing lines or into homologous pap2 plants (T-DNA lines) to generate complementation lines. Through 2 generations of selection on MS agar plate with 50 mg/l Basta (Riedel-deHaen), homologous 35S:PAP2 transgenic lines were obtained. The resistant plants were transferred to soil to grow to maturity, and their transgenic status was further confirmed by PCR and immunoblot analyses. As shown in FIG. **6**A, AtPAP2 protein was overexpressed in OE lines but was absence from the T-DNA line. The homozygous T3 seeds of the transgenic plants were 20 used for further analysis.

To create transgenic AtPAP15 overexpression lines, the cDNA of AtPAP15 was also amplified by RT-PCR and then subcloned into a plant binary vector which bared a kanamy-cin-resistant gene and a cauliflower mosaic virus 35S promoter (CaMV). This expression construct named was then mobilized into *Agrobacterium tumefaciens* strain EHA105 by freeze-thaw transformation (Hofgen and Willmitzer, 1988) and transformed into *Arabidopsis*. Transgenic status was further confirmed by PCR and immunoblot analyses using an anti-AtPAP15 antiserum. As shown in FIG. 6B, AtPAP15 protein was overexpressed in OE lines. The homozygous T3 seeds of the transgenic plants were used for further analysis.

#### 6.3 Production of PAP2 Polycolonal Antiserum and Western Blots Analysis

A fragment of AtPAP2 cDNA corresponding to the N terminal 120 amino acids (from 21 to 141) was amplified 40 using forward primer P2AF (5'-GGTTGAGCTCGAT-TCTAAAGCGACCATTTC-3', SEQ ID NO: 16) and reverse primer P2AR (5'-TTTTGGTACCTCAGGATC-CGAA AGTCAGC-3', SEQ ID NO: 17). The PCR product was cleaved by SacI and KpnI and cloned into the pRsetA 45 vector (Invitrogen) so that the coding sequence of the first 120 a.a. of AtPAP2 was fused to a His-tag sequence. The resulting plasmid was transformed into Escherichia coli strain BL21 (DE3). The BL21 cells were induced at 30° C. by 0.1 mM isopropylthio- $\beta$ -D-galactoside for 4 h and resuspended in 100 mM NaCl and 50 mM Tris-HCl, pH 7.5, 2 mM phenylmethylsulphonyl fluoride (PMSF). The lysates were sonicated 5 times for 30 s each. The overexpressed His-AtPAP2 fusion proteins in inclusion bodies were centrifuged at 5000×g for 15 min, and the pellets were solubi- 55 lized in 150 mM NaCl, 8 M urea, and 20 mM Tris-HCl, pH 7.5. The fusion proteins were purified on a HisTrap FF (GE Healthcare) column and were used for standard immunization protocols in rabbits.

#### 6.4 Expression Analysis of AtPAP2 mRNA and its Protein Levels

The mRNA expression level of AtPAP2 was analyzed by the Spot History program (http://affymetrix.arabidopsisinfo/ 65 narrays/spothistory.pl) that presented the expression levels of a given gene in thousands of microarray (Affymetrix

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ATH1 microarray) database. Spot history analysis indicated that the expression of AtPAP2 was constitutive but is relative low in most experimental circumstances (FIG. 7a). To determine AtPAP2 expression levels, different tissues of wild-type *A. thaliana* (Col-0) were collected.

The expression level of proteins were also studied by western blotting, using the anti-AtPAP2 antiserum generated from Section 6.3. Total plant soluble protein was extracted from wild-type A. thaliana, T-DNA line, AtPAP2-overexpress lines in grinding buffer (Tris-HCl 50 mM, pH7.4 containing 150 mM NaCl, 1 mM EDTA, 0.2 mM PMSF) on ice. Protein extracts were centrifuged at 16000×g and supernatants were collected for Bradford protein concentration determination assay. Equal amount of protein samples (50-90 µg/lane in different experiments) were loaded and separated in 12% (w/v) SDS-PAGE. The separated proteins were transferred to Hybond C-Extra membranes (Amersham Biosciences) (400 mA, 1 h). Membranes were blocked with 5% (w/v) non-fat milk in TTBS washing buffer (pH 7.6) for 2 hours and probed with specific anti-AtPAP2 antiserum for 3 hours or overnight at an 1:1000 dilution at 4° C. After rinsing the membrane with three changes of TTBS washing buffer (20 mM Tris-HCl, pH7.6, 136 mM NaCl, 0.1% Tween20) in half an hour, HRP-labeled secondary antibody, diluted 1:10,000 in TTBS washing buffer was added. After 2 hours, the membrane was washed thrice before the bands were visualized by Enhanced Chemiluminescence method (Amersham Biosciences). As shown in FIG. 7b, AtPAP2 protein was expressed in all tissues tested (Leaf, Flower, Stem, Root, Silique) at equal levels. The protein expression 35 level of AtPAP2 during germination was very stable too (FIG. 7c) and was independent of phosphorus status (FIG.

## 6.5 Growth Phenotypes of WT, T-DNA Line and OE Lines

Arabidopsis seeds were soaked in water at 4° C. for 3 days. The seeds were surface sterilized and sown on Murashige and Skoog (MS) medium supplemented with 2% (w/v) sucrose for 10 days. Seedlings with 2 rosette leaves of the same size were transferred to soil under Long Day (16 h light at 22° C./8 h dark at 18° C.) or Short Day (8 h light at 22° C./16 h dark at 18° C.) conditions in a plant growth chamber. Flowering time was started to be measured by scoring the number of rosette leaves and cauline leaves when the primary inflorescence florescence reached 1 cm above the rosette leaves. Ten to 20 plants were scored for each line (Liu et al., 2008; Wu et al., 2008).

The inflorescences of OE lines of AtPAP2 emerged earlier (5-6 days for Long Day, 14-16 days for Short Day) than that of the WT and T-DNA lines (Table 1). Under Long Day conditioning, the number of rosette leaves of the OE lines were less (5-6 leaves) than the WT during the emergence of inflorescence (Table 1 and FIG. 8). At day 28 (Long Day), the OE lines of AtPAP2 had more cauline leaves and inflorescences than the WT and T-DNA lines, but had less rosette leaves (Table 2.). This phenotype observation was repeated at least four times and the results of one of the experiments were shown here.

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6.6 Growth Phenotypes of Truncated AtPAP2
Constructs

A	AtPAP2 OE lines flowered at an earlier developmental stage.							
	Long Day (16 h/8 h)			Short Day (8 h/16 h)			)	
Lines	AEI	$^{\mathrm{SD}}$	NRL	SD	AEI	SD	NRL	SD
Col-0 T-DNA OE7 OE21	26.9 25.7 20.0* 20.8*	1.2 0.7 1.1 0.6	13.0 11.6 6.4* 6.5*	0.8 1.1 0.5 0.7	41.0 40.7 25.6* 26.0*	4.7 4.9 1.3 1.1	18.0 15.0 5.3* 5.4*	3.0 3.0 0.5 0.5

AEI: Average date of emergence of inflorescence

NRL: No. of rosette leaves at the first appearance of inflorescence

TABLE 2

	Phenotypes of AtPAP2 OE lines at Day 28 (Long Day).							
Lines	No. of Rosette Leaf	SD	No. of Cauline Leaf	SD	No. of Inflorescence	SD		
Col-0 T-DNA OE7 OE21	14.5 16.7 9.9* 10.2*	1.2 1.7 1.0 1.8	1.6 1.9 6.0* 7.2*	0.5 0.6 1.2 1.6	1.0 1.0 3.6* 3.7*	0.0 0.0 0.7 1.1		

<sup>\*</sup>Statistically (p < 0.001) different from the wild-type (n = 15).

At maturity (Long Day), the number of siliques and the total weight of seeds harvested from each line were recorded. Two separate experimental trials are shown in 30 Tables 3A and 3B. Our results showed that overexpression of AtPAP2 resulted in increase number of siliques per plant and the seed yield per plant. Compared to that of the wild-type, the seed yield of the two overexpression lines shown in Table 3A increased 38-40%. Compared to that of 35 the wild-type, the seed yield of the two overexpression lines shown in Table 3B increased 54-58%.

TABLE 3A

	OE lines produced more siliques and seeds (Trial 1).						
Lines	No. of siliques/plant	SD	Weight of seeds (g)/plant	SD	N		
Col-0	327.4	53.3	0.188	0.047	5		
T-DNA	236.6*	60.2	0.121*	0.040	7		
OE7	453.2**	62.1	0.264**	0.039	5		
OE21	498.2#	52.5	0.260**	0.049	7		

Statistically (p < 0.02\*, p < 0.01\*\*, p < 0.001\*\*) different from the wild-type.

TABLE 3B

OE lines produced more siliques and seeds (Trial 2).						
Lines	No. of siliques/plant	SD	Weight of seeds (g)/plant	SD	N	
Col-0	396.4	89.5	0.225	0.058	13	
T-DNA	386.3	70.4	0.240	0.049	12	
OE7	610.9*	76.6	0.351*	0.050	7	
OE21	624.9*	94.7	0.355*	0.066	11	

Statistically (p < 0.0001\*) different from the wild-type.

However, the OE lines of AtPAP15 grew normally and were not different from the wild-types. Therefore, the enhanced growth performance was due to the overexpression of AtPAP2, which bears a transmembrane-like motif at its C-terminus (FIGS. 2 and 3).

An alternate vector construct employing the sequence for AtPAP2 was also constructed using analogous techniques to those described above. As shown in FIG. 12, a construct equivalent to the OE lines of AtPAP2 missing the C-terminal motif (residues 614-636 of SEQ ID NO: 2) was constructed (P2C lines). Transgenic plants were generated using substantially identical techniques to those described above. Western blot analysis was used to confirm the over-expression of the AtPAP2 fragment proteins in transformed plant lines. Performance of Western blot analysis was identical to that reported above. As show in FIG. 13, the P2C lines were strongly overexpressed. The growth phenotype of the P2C lines appeared to be indifferent from the wild-type, which is indicative of the importance of the C-terminal domain of AtPAP2 in developing an increased growth phenotype.

#### 6.7 MS/MS Analysis of Sucrose and Glucose Levels in Leaf

Rosette leaves of plants of various developmental stages were harvested at the end of the light period of 21-day-old plants. Soluble sugars were extracted from Arabidopsis using chloroform/methanol method (Lunn et al., 2006; Antonio et al., 2007; Luo et al., 2007). 100 mg plant tissues were ground to a fine powder in liquid nitrogen and mixed and vortexed with 250 µl ice-cold chloroform: methanol (3:7, v/v). Soluble metabolites were then extracted at -20° C. overnight. 200 µl water was added to the mixture with repeated shaking. The extracts were centrifuged at 16000xg for 10 min and the supernatant was collected. The pellet was re-extracted by 200 µl water and the supernatant was collected by centrifugation as described above. The combined supernatant was evaporated to dryness using a SpeedVac and the pellet was re-dissolved in 200 µl water. Finally, debris was removed by centrifugation at 16000×g for 30 min.

20 µl filtered samples were analyzed by an API-3000 40 triple-quadrupole mass spectrometer (Applied Biosystems) via an electrospray ionization source. The parameters, optimized by 0-40 μg/ml glucose and sucrose standards, were as following: curtain gas (CUR) 25, nebulizer gas (GS1) 50, auxiliary gas (GS2) 30, ionspray voltage -4.5 k V, temperature 400° C., declustering potential (DP) -106 V, entrance potential (EP) -8.5 V, collision cell entrance potential (CEP) -46.7 V, collision energy (CE) 20 V. The peaks were identified by comparison with glucose and sucrose standards and the amount of sugars were quantified by standard curves 50 of these sugars. The Analyst 1.3.1 software (Applied Biosystems) was used for data acquisition, peak integration, and calculation. The amount of sucrose and glucose at the end of day in the shoots of 21-day-old soil grown plants were shown in FIG. 9. It was found that the levels of both sugars 55 were significantly higher than that in WT.

#### 6.8 Recovery of Plants after Prolonged Darkness Treatment

Seeds of wild-type, T-DNA, OE7 and OE21 lines were germinated in MS (2% sucrose) medium for 10 days. Seedlings with 2 small visible rosette leaves (1 mm) of the same size were transferred to soil for another 12 days in normal growth conditions (LDs, 16 h/light (22° C.)/8 h darkness (18° C.)). The light source of the growth chamber was then switched off for 12 days. Then the plants were allowed to recover under the 16 h/light (22° C.)/8 h darkness

<sup>\*</sup>Statistically (p < 0.001) different from the wild-type (n = 15).

 $(18^{\circ}\ \text{C.})$  cycle for 10 days. The plants that stayed green and that continued to emerge inflorescence were recorded in Table 4.

TABLE 4

Surviving rate and flowering ratio after prolonged darkness treatment.				
	Flowering after recovery	Recovery (leaf greening)		
Wild-type	8/12	11/12		
T-DNA	5/12	8/12		
OE7	9/9	9/9		
OE21	12/12	12/12		

Extended darkness could induce carbohydrate starvation (Thompson et al., 2005). Our data showed that the OE lines exhibited 100% recovery rate under prolonged (12 days) darkness treatment, which was higher than that of the WT and the T-DNA line (FIG. 10). This could be attributed to a higher endogenous sugar levels (FIG. 9) in the OE lines.

#### 6.9 Phenotypes of Plants Under NaCl and ABA Treatments

5-day-old seedlings grown on MS agar were transferred to MS agar with NaCl (50 mM, 100 mM, 150 mM), ABA (0.1 uM, 0.2 uM. 0.5 uM, 1 uM, 2 uM) or sorbitol (300 mM, 400 mM, 500 mM). Alternatively, seeds were directly germinated on the treatment media. Wild-type, T-DNA and OE 30 lines did not show remarkable phenotypic differences under the above conditions.

#### 6.10 Subcellular Fractionation

Rosette leaves of three-week-old wild-type (Col-0) Arabidopsis were harvested and stored at -80° C. freezer until use. Tissue (4-5 g) were ground to fine powder in liquid nitrogen using a mortar with a pestle. The powder was transferred into 10 ml grinding buffer (0.3 M sucrose, 40 40 mM Tris-HCl (pH 7.8), 5 mM MgCl<sub>2</sub>, 1 mM PMSF) and swelled on ice for 5 min. Homogenization was performed for two 30-second pulses at high-speed setting. The homogenate was filtered through two layers of Miracloth (Tetko, Elmsford, N.Y., USA). Subsequently, the homogenate was 45 separated by centrifugation at 350 g for 10 min at 4° C. The pellet (crude nuclear) was further layered onto 1 ml of 2.3 M sucrose, 50 mM Tris-HCl (pH 8.8), 5 mM MgCl<sub>2</sub> in an Eppendorf tube for centrifugation at 15,000 g 10 min at 4° C., to obtain the nuclear fraction in the derived pellet. 50 Supernatants from the first low-speed centrifugation (350 g) were centrifuged at 12,000×g for 20 min at 4° C. The pellet contained large particles including mitochondria, chloroplasts and peroxisomes. The supernatant was further centrifuged at 100,000×g for 1 h at 4° C. to yield the soluble 55 cytosol fraction in the resulting supernatant. The pellet representing the membrane fraction was resuspended in 0.1 ml grinding buffer. Protein concentration in the extract was determined following the method of Bradford (Bradford, 1976) using the Bio-Rad Protein Assay Kit I.

To isolate cell wall, leaf tissues were homogenized in grinding buffer (62.5 mM Tris-HCl, pH 7.5, 5 mM MT, 1% (v/v) bovine serum albumin, 2 mM phenylmethylsulfonyl fluoride, 2  $\mu$ g/ml leupeptin, 2  $\mu$ g/ml E-64, 2  $\mu$ g/ml pepstatin A) using a Polytron (full speed, 3×10 s). The homogenate 65 was centrifuged at 1,000×g for 3 min. The pellet was washed with ice-cold grinding buffer (without 1% BSA) 10 times.

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Finally the (cell wall) pellet was washed by resuspending in 500 mM CaCl<sub>2</sub>, 20 mM NaCl, 62.5 mM Tris-HCl, pH 7.5, and spinning at 10,000×g for 15 min (He et al., 1996).

The subcellular fractions were run in a SDS-PAGE gel and were probed with anti-AtPAP2 antiserum. AtPAP2 was detected in membrane and soluble protein fractions but not in nucleus, mitochondria nor chloroplasts (FIG. 11).

In summary, *Arabidopsis* plants transformed with the AtPAP2 gene have the following phenotypes when they were compared with the wild-type: (1) Faster growth rate (Tables 1 and 2); (2) Higher sucrose content (FIG. 9); (3) Higher glucose content (FIG. 9); and (4) Higher crop yield (Table 3).

Those skilled in the art will recognize, or be able to ascertain many equivalents to the specific embodiments described herein using no more than routine experimentation. Such equivalents are intended to be encompassed by the following claims.

With respect to any figure or numerical range for a given characteristic, a figure or a parameter from one range may be combined with another figure or a parameter from a different range for the same characteristic to generate a numerical range.

All publications, patents and patent applications mentioned in this specification are herein incorporated by reference into the specification to the same extent as if each individual publication, patent or patent application was specifically and individually indicated to be incorporated herein by reference.

Citation or discussion of a reference herein shall not be construed as an admission that such is prior art to the present patent application.

While the embodiments have been explained in relation to certain embodiments, it is to be understood that various modifications thereof will become apparent to those skilled in the art upon reading the specification. Therefore, it is to be understood that the embodiments disclosed herein are intended to cover such modifications as fall within the scope of the appended claims. Features of two or more of any of the above embodiments can be combined to form additional embodiments.

Other than in the operating examples, or where otherwise indicated, all numbers, values and/or expressions referring to quantities of ingredients, reaction conditions, etc., used in the specification and claims are to be understood as modified in all instances by the term "about."

### 6.11 Assays of enzymes involved in sucrose metabolism

Sucrose phosphate synthesis (SPS), sucrose synthesis (SuSy), cytosolic invertase and cell wall invertase activities in the shoot of 20-day-old plants were determined. Samples were collected 8 h after the light and dark period (Long Day). SPS activity was measured under both optimal (Vmax) and limiting (V limit) assay conditions (Park et al., 2008). SuSy, cytosolic invertase and insoluble cell wall invertase activities were also determined (Doehlert, 1987). The assays were repeated three times and the SPS (Vmax and V limit) activities of both independent lines were significantly higher than that of the wild-type and T-DNA lines in all three repeated experiments. The data of a representative experiment is shown in table 5. In contrast to SPS, SuSy, cytosolic invertase and cell wall invertase activities were not different among the lines.

Enzyme assays							
Plant line	e WT	T-DNA	OE7	OE21			
Sucr	Sucrose phosphate synthase (µM sucrose/µg enzyme extracts/hour)						
Vmax (Day)	108.8 ± 18.1	116.1 ± 11.9	157.9 ± 21.7**	159.5 ± 19.0**			
Vlimit (Day)	63.6 ± 6.4	69.5 ± 2.5	90.8 ± 18.5**	79.8 ± 13.7*			
Vmax (Night)	118.3 ± 11.4	104.9 ± 14.4	150.9 ± 19.4**	136.5 ± 15.1*			
Vlimit (Night)	$74.9 \pm 6.3$	$56.7 \pm 3.3$	93.4 ± 3.6**	97.3 ± 10.9**			
( 3)	Sucrose synthas	e (uM glucose/	ug enzyme extra	cts/hour)			

#### $255.2 \pm 5.4$ $249.2 \pm 4.6$ 247.0 ± 24.0 248.6 ± 4.5 Day 249.4 ± 7.2 $252.7 \pm 8.8$ $250.8 \pm 8.7$ $258.8 \pm 5.5$ Night Cytosolic invertase ( $\mu M$ glucose/ $\mu g$ enzyme extracts/hour) $14.3 \pm 2.3$ $12.1 \pm 5.4$ $18.3 \pm 9.1$ $18.0 \pm 0.8$ Day (Acid) $26.1 \pm 10.9$ $17.6 \pm 4.3$ $13.8 \pm 0.8$ $14.0 \pm 6.8$ Night (Acid) $169.7 \pm 9.8$ 161.2 ± 32.3 160.9 ± 27.9 $178.8 \pm 16.9$ Day (Alkaline) $130.0 \pm 8.2$ $105.1 \pm 12.6 \quad 136.1 \pm 13.6$ $136.6 \pm 1.4$ Night (Alkaline) Cell wall invertase (µM sucrose/µg enzyme extracts/hour)

Day (Acid)	22.3 ± 3.9	18.0 ± 4.9	23.5 ± 6.3	$24.0 \pm 6.6$
Night (Acid)	21.6 ± 10.1	$17.0 \pm 4.3$	$28.3 \pm 4.2$	$22.3 \pm 3.3$
Day (Alkaline)	121.7 ± 2.8	127.1 ± 2.2	$101.8 \pm 6.4$	$105.5 \pm 2.0$
Night (Alkaline)	138.9 ± 6.3	151.5 ± 8.7	123.4 ± 5.6	135.4 ± 14.7

(\*\*P < 0.01; \*P < 0.05)

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Met Ile Pro Asp Leu Pro Leu Pro Phe Leu Phe Ser Leu Phe Ile Ile

Phe Phe His Leu Ala Glu Ser Lys Pro Ser Leu Thr Ala Thr Pro Thr 20 25 30

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<sup>&</sup>lt;213> ORGANISM: Glycine max

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PIO	Ser 50	PIO	ser	Asp	ьец	55	Pile	ьец	AIA	TTE	60	ser	PIO	PIO	1111
Ser 65	Pro	His	Asp	Asn	Phe 70	Ile	Gly	Tyr	Leu	Phe 75	Leu	Ser	Gln	Ser	Ala 80
Thr	Trp	Arg	Thr	Gly 85	Ser	Gly	Asn	Leu	Ser 90	Leu	Pro	Leu	Val	Asp 95	Leu
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Tyr	Asn	Thr 275	Phe	Leu	Arg	Thr	Gln 280	Asp	Glu	Ser	Ile	Ser 285	Thr	Met	Lys
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Met Leu Glu His Leu Glu Pro Leu Leu Val Asn Asn Asn Val Thr Leu Ala Leu Trp Gly His Val His Arg Tyr Glu Arg Phe Cys Pro Leu Asn Asn Phe Thr Cys Gly Val Asn Ala Gly His Asn Ala Gly Asp Lys Lys Gly Tyr Thr Val His Ile Val Ile Gly Met Ala Gly Gln Asp Trp Gln 520 Pro Val Trp Glu Pro Arg Pro Asp His Pro Asp Asp Pro Ile Phe Pro Gln Pro Lys Trp Ser Leu Tyr Arg Gly Gly Glu Phe Gly Tyr Thr Arg Leu Val Ala Thr Lys Gln Lys Leu Val Leu Ser Tyr Val Gly Asn His Asp Gly Glu Val His Asp Gln Leu Glu Ile Leu Ala Ser Gly Glu Val 585 Val Ser Gly Asp Gly Gly Cys Ser Ile Ala Asp Ala Asn Ser Lys Ala 600 Gly Asn Val Ile Val Glu Ser Thr Leu Ser Trp Tyr Val Lys Gly Gly 615 Ser Val Leu Leu Gly Ala Phe Met Gly Tyr Val Phe Gly Tyr Val 630 635 Thr Ser Ala Arg Lys Lys Ser Glu Val Pro Glu Ser Asn Trp Thr Pro 650 Val Lys Thr Glu Glu Thr 660 <210> SEQ ID NO 7 <211> LENGTH: 1965 <212> TYPE: DNA <213 > ORGANISM: Zea mays <400> SEQUENCE: 7 atgtaccccg aaaaccccca cctccgcttc ctcctcttcc tcgccgtcgc ggcagttgcc 60 gccggcgggg ctgcggcgaa caccaccctc accgcgtccc tctccggcaa ccagatcaag 120 atcatctggt ccggactccc ggccccggac ggcctcgact acgttgccat ctactcgccg cogtoctocc togacogoga cttoctoggo tatotottoc toaacggoto cgcotoctgg cgcggcggct ccggggagct ctccctcccg ctcctcccga cgctccgcgc gccctaccag ttccgtctct ttcgctggcc cgccaaggag tactcctacc accacgtcga ccacgaccag aaccegetee eecaeggeaa geacegegte geegteteeg eegaegtete egteggegae cccgcccgcc ccgagcagct gcacctcgcg tttgcggatg aggtcgacga gatgcgggtc 480 540 ctgttcgtgt gcggcgaccg cggggagagg gtcgtcaggt acgggctgca gaaggaggac gacaaggagt ggaaggaggt gggcacggat gtgagcacgt acgagcagag gcacatgtgc 600 gattggccgg ccaacagcag cgtcgcctgg agggatccgg gattcgtctt cgacggcctc atgaagggat tggagcccgg aaggaggtac ttttacaagg ttggtagtga cacaggagga 720 tggagtgaga tatacagctt catttcacgt gacagtgaag ccagtgagac caatgctttt 780 ctatttqqtq acatqqqaac ttatqtqcct tataacacct acattcqcac acaatctqaq 840 agettgteca etgtaaagtg gateettegt gatattgaag eeettggaga taaaeeegee 900

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<213> ORGANISM: Zea mays

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Ser Leu Ser Gly Asn Gln Ile Lys Ile Ile Trp Ser Gly Leu Pro Ala 35  $\phantom{\bigg|}40\phantom{\bigg|}40\phantom{\bigg|}$ 

Pro Asp Gly Leu Asp Tyr Val Ala Ile Tyr Ser Pro Pro Ser Ser Leu 50 55 60

Asp Arg Asp Phe Leu Gly Tyr Leu Phe Leu Asn Gly Ser Ala Ser Trp 65 70 75 80

Arg Gly Gly Ser Gly Glu Leu Ser Leu Pro Leu Leu Pro Thr Leu Arg

Tyr His His Val Asp His Asp Gln Asn Pro Leu Pro His Gly Lys His 115 \$120\$

Arg Val Ala Val Ser Ala Asp Val Ser Val Gly Asp Pro Ala Arg Pro

Glu Gln Leu His Leu Ala Phe Ala Asp Glu Val Asp Glu Met Arg Val

Leu Phe Val Cys Gly Asp Arg Gly Glu Arg Val Val Arg Tyr Gly Leu

Gln Lys Glu Asp Asp Lys Glu Trp Lys Glu Val Gly Thr Asp Val Ser \$180\$

Thr	Tyr	Glu 195	Gln	Arg	His	Met	Cys 200	Asp	Trp	Pro	Ala	Asn 205	Ser	Ser	Val
Ala	Trp 210	Arg	Asp	Pro	Gly	Phe 215	Val	Phe	Asp	Gly	Leu 220	Met	ГÀа	Gly	Leu
Glu 225	Pro	Gly	Arg	Arg	Tyr 230	Phe	Tyr	Lys	Val	Gly 235	Ser	Asp	Thr	Gly	Gly 240
Trp	Ser	Glu	Ile	Tyr 245	Ser	Phe	Ile	Ser	Arg 250	Asp	Ser	Glu	Ala	Ser 255	Glu
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Leu	Arg 290	Asp	Ile	Glu	Ala	Leu 295	Gly	Asp	Lys	Pro	Ala 300	Phe	Ile	Ser	His
Ile 305	Gly	Asp	Ile	Ser	Tyr 310	Ala	Arg	Gly	Tyr	Ser 315	Trp	Val	Trp	Tyr	His 320
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Asp	Met	Val	Glu 580	Ile	Phe	Ser	Gly	Leu 585	Val	Ser	Pro	Ser	Asn 590	Ser	Ser
Val	Ala	Glu 595	Ala	Val	Asp	Gly	Thr 600	Lys	Leu	Gly	Thr	Gly 605	Val	Ser	Thr
Val	Arg	Lys	Ile	Ser	Pro	Leu	Tyr	Leu	Glu	Ile	Gly	Gly	Ser	Val	Met

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610
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Ser Arg Se 35	r Gly Asp S	er Ile Leu 40	Ile Lys Trp	Ser Asn Leu Asp Ser 45	
Pro Ser As	p Leu Asp T	rp Leu Gly 55	Ile Tyr Ser	Pro Pro Ala Ser Pro 60	
His Asp Hi 65	s Phe Ile G 7		Phe Leu Asn 75	Ala Ser Pro Thr Trp 80	
Gln Ser Gl	y Ser Gly A 85	la Ile Ser	Leu Pro Leu 90	Thr Asn Leu Arg Ser 95	
Asn Tyr Th	r Phe Arg I 100		Trp Thr Gln 105	Ser Glu Ile Asn Pro 110	
Lys His Ly 11		sp Gln Asn 120	Pro Leu Pro	Gly Thr Lys His Leu 125	
Leu Ala Gl	u Ser Glu G	ln Val Gly 135	Phe Gly Ser	Ala Gly Val Gly Arg 140	
Pro Glu Gl:		eu Ala Phe 50	Glu Asp Lys 155	Val Asn Arg Met Gln 160	
Val Thr Ph	e Val Ala G 165	ly Asp Gly	Glu Glu Arg 170	Phe Val Arg Tyr Gly	
Glu Ala Gl	u Asp Ala L 180		Ser Ala Ala 185	Ala Arg Gly Ile Arg 190	

Tyr Glu Arg Glu His Met Cys Asn Ala Pro Ala Asn Ser Thr Val Gly 195 200 205

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Leu 465	Glu	Pro	Leu	Phe	Val 470	Glu	Asn	Asn	Val	Thr 475	Leu	Ala	Leu	Trp	Gly 480
His	Val	His	Arg	Tyr 485	Glu	Arg	Phe	Сла	Pro 490	Ile	Ser	Asn	Asn	Thr 495	CAa
Gly	ГЛа	Gln	Trp 500		Gly	Ser	Pro	Val 505		Leu	Val	Ile	Gly 510	Met	Gly
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Phe	Val	Gly	Asn	His 565	Asp	Gly	Glu	Val	His 570	Asp	Ser	Val	Glu	Met 575	Phe
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Trp	Tyr 610	Val	Lys	Gly	Ala	Gly 615	Leu	Leu	Val	Ile	Gly 620	Val	Leu	Leu	Gly
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1860

1920 1965

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Val	Gly 210	Trp	Arg	Asp	Pro	Gly 215	Phe	Val	Phe	Asp	Gly 220	Leu	Met	Asn	Gly
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		Ile	260					265					270		
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His	Phe	Phe	Ser	Gln 325	Ile	Glu	Pro	Ile	Ala 330	Ala	Asn	Thr	Pro	Tyr 335	His
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Pro	Ser	Trp 355	Ser	Thr	Tyr	Gly	360	Asp	Gly	Gly	Gly	Glu 365	Сув	Gly	Ile
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Gly	Ser	Asp	Gln 420	His	Asn	Phe	Leu	Lys 425	Ala	Asp	Leu	Glu	Lys 430	Val	Asn	
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Phe 625	Ala	Leu	Met	Leu	Gly 630	Phe	Ala	Leu	Gly	Phe 635	Leu	Leu	Arg	Lys	Lys 640	
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Thr Leu Thi	: Lys Ser G	ly Asp Thr 40	Val Thr Let	Arg Trp Sei 45	Gly Ile	
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Arg Ser Asr	n Tyr Ser P 100	he Arg Ile	Phe His Trp	Ser Gln Ser		

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		435			Val		440					445			
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77 78

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Ile	His	Leu	Ala	Leu 165	Ser	Ser	Asp	Glu	Thr 170	Ala	Val	Arg	Val	Met 175	Phe		
Val	Thr	Arg	Asp 180	Pro	Leu	Arg	Ser	Gln 185	Val	Arg	Phe	Gly	Glu 190	Asp	Gly		
Asp	Glu	Leu 195	Gly	Asn	Thr	Val	Asp 200	Ala	Thr	Ser	Val	Thr 205	Tyr	Ser	Gln		
Ile	Asp 210	Met	СЛа	Asp	Glu	Pro 215	Ala	Ser	Ser	Tyr	Gly 220	Trp	Arg	Ser	Pro		
Gly 225	Tyr	Ile	His	Asn	Val 230	Val	Met	Gly	Gly	Leu 235	Asn	Pro	Gly	Ser	Arg 240		
Tyr	Phe	Tyr	Arg	Val 245	Gly	Ser	Asn	Val	Gly 250	Gly	Trp	Ser	Ser	Thr 255	Tyr		
Ser	Phe	Ile	Ala 260	Pro	His	Pro	Arg	Ala 265	Asp	Glu	Thr	Asn	Ala 270	Leu	Ile		

Phe Gly Asp Met Gly Thr Ser Ile Pro Tyr Ser Thr Tyr Gln Tyr Thr

		275					200					205			
		275					280					285			
Gln	Ser 290	Glu	Ser	ràa	Asn	Thr 295	Val	Lys	Trp	Leu	Thr 300	Arg	Asp	Leu	Glu
Gln 305	Ile	Gly	Asp	Lys	Pro 310	Ser	Phe	Val	Ala	His 315	Ile	Gly	Asp	Ile	Ser 320
Tyr	Ala	Arg	Gly	Leu 325	Ser	Trp	Leu	Trp	Asp 330	Asn	Phe	Phe	Thr	Gln 335	Ile
Glu	Pro	Val	Ala 340	Ala	Arg	Ser	Pro	Tyr 345	His	Val	Cys	Met	Gly 350	Asn	His
Glu	Tyr	Asp	Trp	Pro	Gly	Gln	Pro 360	Phe	Lys	Pro	Asp	Trp 365	Ser	Pro	Tyr
Gln	Thr 370	Asp	Gly	Gly	Gly	Glu 375	Cys	Gly	Val	Pro	Tyr 380	Ser	Leu	Arg	Phe
Ile 385	Met	Pro	Gly	Asn	Ser 390	Ser	Leu	Pro	Thr	Gly 395	Thr	Thr	Ser	Pro	Ala 400
Thr	Lys	Asn	Leu	Tyr 405	Tyr	Ser	Ile	Asp	Val 410	Gly	Val	Val	His	Phe 415	Leu
Phe	Tyr	Ser	Thr 420	Glu	Thr	Asp	Phe	Gln 425	Val	Gly	Ser	Pro	Gln 430	Tyr	Thr
Phe	Ile	Ala 435	Asn	Asp	Leu	Arg	Thr 440	Val	Asp	Arg	Asn	Lуs 445	Thr	Pro	Phe
Val	Val 450	Phe	Leu	Gly	His	Arg 455	Pro	Leu	Tyr	Thr	Thr 460	Asp	Tyr	Arg	Ala
Leu 465	Leu	Asp	Thr	Met	Thr 470	Gln	Lys	Leu	Val	Gln 475	Thr	Phe	Glu	Pro	Leu 480
Leu	Ile	Asp	Thr	Asn 485	Val	Thr	Val	Ala	Phe 490	Сув	Gly	His	Val	His 495	Lys
Tyr	Glu	Arg	Met 500	Сув	Pro	Leu	Lys	Asn 505	Tyr	Thr	Cys	Ile	Glu 510	Pro	Ser
Lys	Ala	Asn 515	Gly	Glu	Leu	Pro	Ile 520	His	Met	Val	Val	Gly 525	Met	Gly	Gly
Ala	Asp 530	His	Gln	Pro	Ile	Asp 535	Asp	Pro	Leu	Pro	Ser 540	Gln	Ser	Gln	Pro
Ile 545	Phe	Pro	Gln	Pro	Ser 550	Trp	Ser	Val	Phe	Arg 555	Thr	Phe	Glu	Trp	Gly 560
Tyr	Ile	Arg	Leu	His 565	Ala	Thr	Arg	His	Leu 570	Met	Thr	Ile	Ser	Tyr 575	Val
Gly	Asn	His	Asp 580	Gly	Lys	Val	His	Asp 585	Val	Val	Glu	Ile	Pro 590	Val	Leu
Asp	Asp	Ile 595	Lys	Ser	Gly	Ala	Tyr 600	Val	Glu	Ser	Arg	Glu 605	Ser	Phe	Phe
Asp	Thr 610	Ala	Ser	Gly	Val	Gln 615	Ile	Pro	Cys	Gly	Arg 620	Ser	Glu	Asn	Ile
Val 625	Ala	Phe	Leu	Phe	Val 630	Leu	Ala	Leu	Gly	Сув 635	Gly	Cys	Gly	Ala	Ala 640
Ala	Thr	Leu	Phe	Phe 645	Met	Arg	Arg	Gln	Gln 650	Arg	Lys	Gln	Ile	Trp 655	Gln
Pro	Val	Asn	Arg 660	Glu	Glu	Ala	Ser	Ser 665	Ser	Gln	Leu				

<sup>&</sup>lt;210> SEQ ID NO 26 <211> LENGTH: 1944 <212> TYPE: DNA <213> ORGANISM: Populus trichocarpa

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atttcctggt ccaacgttga ttcaccttcc aaactcgact ggctcgggct ctattcacct
                                                                     180
cetgacteae etcaegacea etteattgge tacaagttee tttetteete teetteatgg
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                                                                    1200
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                                                                    1260
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                                                                    1320
gggcacagac caatgtatac tactagcaat gaaaacaggg atgccccaat gaggaacaaa
                                                                    1380
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<210> SEQ ID NO 27
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Met Lys Leu Pro Ile Phe Leu Leu Leu Leu Leu Leu Ser Leu Ile Thr

<sup>&</sup>lt;211> LENGTH: 647

<sup>&</sup>lt;212> TYPE: PRT

<sup>&</sup>lt;213> ORGANISM: Populus trichocarpa

<sup>&</sup>lt;400> SEQUENCE: 27

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Gln	Lys	Ser 35	Gly	Asp	Thr	Val	Thr 40	Ile	Ser	Trp	Ser	Asn 45	Val	Asp	Ser
Pro	Ser 50	Lys	Leu	Asp	Trp	Leu 55	Gly	Leu	Tyr	Ser	Pro 60	Pro	Asp	Ser	Pro
His 65	Asp	His	Phe	Ile	Gly 70	Tyr	Lys	Phe	Leu	Ser 75	Ser	Ser	Pro	Ser	Trp 80
Gln	Ser	Gly	Ser	Gly 85	Ser	Ile	Ser	Leu	Pro 90	Ile	Thr	Asn	Leu	Arg 95	Ser
Asn	Tyr	Ser	Phe 100	Arg	Ile	Phe	His	Trp 105	Thr	Glu	Ser	Glu	Ile 110	Asn	Pro
ГЛа	Arg	His 115	Asp	His	Asp	His	Asn 120	Pro	Leu	Pro	Gly	Thr 125	Ala	His	Phe
Leu	Ala 130	Glu	Ser	Asp	Val	Val 135	Gly	Phe	Glu	Ser	Gly 140	His	Gly	Pro	Glu
Gln 145	Ile	His	Leu	Ala	Tyr 150	Thr	Asp	Asp	Glu	Asp 155	Glu	Met	Arg	Val	Met 160
Phe	Val	Val	Gly	Asp 165	Gly	Glu	Glu	Arg	Gly 170	Val	Lys	Trp	Gly	Glu 175	Arg
Asp	Gly	Glu	Trp 180	Ser	His	Val	Ser	Gly 185	Ala	Arg	Val	Val	Arg 190	Tyr	Glu
Arg	Glu	Asp 195	Met	Cys	Asp	Ala	Pro 200	Ala	Asn	Gly	Ser	Ile 205	Gly	Trp	Arg
Asp	Pro 210	Gly	Trp	Ile	His	Asp 215	Gly	Val	Met	Lys	Asp 220	Leu	Lys	Lys	Gly
Val 225	Arg	Tyr	Tyr	Tyr	Gln 230	Val	Gly	Ser	Asp	Ser 235	Lys	Gly	Trp	Ser	Thr 240
Thr	Arg	Ser	Phe	Val 245	Ser	Arg	Asn	Gly	Asp 250	Ser	Asp	Glu	Thr	Ile 255	Ala
Phe	Leu	Phe	Gly 260	Asp	Met	Gly	Thr	Ser 265	Thr	Pro	Tyr	Ala	Thr 270	Phe	Ile
Arg	Thr	Gln 275	Aap	Glu	Ser	Ile	Ser 280	Thr	Met	ГЛа	Trp	Ile 285	Leu	Arg	Asp
Ile	Glu 290	Ala	Ile	Gly	Asp	Lys 295	His	Ala	Phe	Val	Ser 300	His	Ile	Gly	Asp
Ile 305	Ser	Tyr	Ala		Gly 310		Ser	Trp		Trp 315		His	Phe	Phe	Thr 320
Gln	Val	Glu	Pro	Val 325	Ala	Ser	Lys	Val	Pro 330	Tyr	His	Val	Сув	Ile 335	Gly
Asn	His	Glu	Tyr 340	Asp	Trp	Pro	Leu	Gln 345	Pro	Trp	Lys	Pro	350	Trp	Ala
Asn	Ala	Val 355	Tyr	Gly	Thr	Asp	Gly 360	Gly	Gly	Glu	Càa	Gly 365	Val	Pro	Tyr
Ser	Leu 370	Lys	Phe	Asn	Met	Pro 375	Gly	Asn	Ser	Ser	Asp 380	Ser	Thr	Gly	Thr
Arg 385	Ala	Pro	Ala	Thr	Arg 390	Asn	Leu	Tyr	Tyr	Ser 395	Phe	Asp	Thr	Gly	Ala 400
Val	His	Phe	Val	Tyr 405	Ile	Ser	Thr	Glu	Thr 410	Asn	Phe	Val	Ala	Gly 415	Ser
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Lys Thr	Pro 435	Phe	Val	Val	Val	Gln 440	Gly	His	Arg	Pro	Met 445	Tyr	Thr	Thr	
Ser Asn 450	Glu	Asn	Arg	Asp	Ala 455	Pro	Met	Arg	Asn	Lys 460	Met	Leu	Glu	His	
Leu Glu 465	Pro	Leu	Phe	Thr 470	Lys	Tyr	Asn	Val	Thr 475	Leu	Ala	Leu	Trp	Gly 480	
His Val	His	Arg	Tyr 485	Glu	Arg	Phe	Cys	Pro 490	Val	Asn	Asn	Phe	Ile 495	Сув	
Gly Ser	Thr	Trp 500	Lys	Gly	Phe	Pro	Val 505	His	Ala	Val	Ile	Gly 510	Met	Ala	
Gly Gln	Asp 515	Trp	Gln	Pro	Ile	Trp 520	Glu	Pro	Arg	Ser	Asp 525	His	Pro	Asn	
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Ala Ser	Gly	Glu 580	Val	Leu	Ser	Gly	Asp 585	Asp	Ser	Ile	Ser	Val 590	Asp	Ala	
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Gly Ala 610	Ser	Val	Leu	Val	Leu 615	Gly	Ala	Phe	Val	Gly 620	Tyr	Thr	Leu	Gly	
Tyr Ala 625	Ser	His	Ser	Arg 630	Lys	Gln	Asn	Gly	Asn 635	Lys	Ala	Ser	Trp	Thr 640	
Pro Val	Lys	Ser	Glu 645	Asp	Ile										
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atccgctg															180
ccgtcctc															240
cgcggcgg															300
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aacccgct															420
cccgcgcg															480
ctgttcgt															540
gagaagga															600
gattggcc															660
atgaaggg															720
tggagtga															780
ctgtttgg															840
	ישר כי		שמשמי		9 '	- 5000	- 541			a su	9'		Jau	,~~ J~ J	515

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900

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<213> ORGANISM: Saccharum officinarum

<400> SEQUENCE: 29

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Thr Leu Ser Ser Asp Gln Ile Lys Ile Arg Trp Thr Gly Leu Pro Ala

Asp Arg Asp Phe Leu Gly Tyr Leu Phe Leu Asn Gly Ser Ala Ser Trp 65 70 75 80

Arg Gly Gly Ser Gly Glu Leu Ser Leu Pro Arg Leu Pro Thr Leu Arg

Ala Pro Tyr Gln Phe Arg Leu Phe Arg Trp Pro Ala As<br/>n Glu Tyr Ser 100 105 110

Tyr His His Ile Asp His Asp Arg Asn Pro Leu Pro His Gly Lys His 120

Arg Val Ala Val Ser Ala Asp Val Ser Val Gly Asp Pro Ala Arg Pro

Glu Gln Val His Leu Ala Phe Ala Asp Gly Ile Asp Glu Met Arg Val

Leu Phe Val Cys Gly Asp Arg Gly Lys Arg Val Val Arg Tyr Gly Leu

Gln Lys Glu Asp Glu Lys Glu Trp Lys Glu Val Gly Thr Asp Val Ser

			180					185					190		
Thr	Tyr	Lys 195	Gln	Lys	His	Met	Cys 200	Asp	Trp	Pro	Pro	Asn 205	Ser	Ser	Val
Ala	Trp 210	Arg	Asp	Pro	Gly	Phe 215	Val	Phe	Asp	Gly	Leu 220	Met	Lys	Gly	Leu
Glu 225	Pro	Gly	Arg	Arg	Tyr 230	Phe	Tyr	Lys	Val	Gly 235	Ser	Asp	Thr	Gly	Gly 240
Trp	Ser	Glu	Ile	Tyr 245	Ser	Phe	Ile	Ser	Arg 250	Asp	Ser	Glu	Ala	Asn 255	Glu
Thr	Asn	Thr	Phe 260	Leu	Phe	Gly	Asp	Met 265	Gly	Thr	Tyr	Val	Pro 270	Tyr	His
Thr	Tyr	Ile 275	Arg	Thr	Gln	Asp	Glu 280	Ser	Leu	Ser	Thr	Val 285	Lys	Trp	Ile
Leu	Arg 290	Asp	Ile	Glu	Ala	Leu 295	Gly	Asp	Lys	Pro	Ala 300	Phe	Ile	Ser	His
Ile 305	Gly	Asp	Ile	Ser	Tyr 310	Ala	Arg	Gly	Tyr	Ser 315	Trp	Val	Trp	Asp	His 320
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Tyr	Ser 370	Val	Lys	Phe	Arg	Met 375	Pro	Gly	Asn	Ser	Ile 380	Leu	Pro	Thr	Gly
Asn 385	Gly	Gly	Pro	Asp	Thr 390	Arg	Asn	Leu	Tyr	Tyr 395	Ser	Phe	Asp	Ser	Gly 400
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Ser	Arg	Thr 435	Pro	Phe	Val	Val	Phe 440	Gln	Gly	His	Arg	Pro 445	Met	Tyr	Thr
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His 465	Leu	Glu	Pro	Leu	Leu 470	Val	Thr	Tyr	Ser	Val 475	Thr	Leu	Ala	Leu	Trp 480
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Cya	Val	Asn	Thr 500	Ser	Ser	Ser	Phe	Gln 505	Tyr	Ser	Gly	Ala	Pro 510	Val	His
Leu	Val	Ile 515	Gly	Met	Gly	Gly	Ala 520	Asp	Trp	Ala	Thr	Ile 525	Trp	Gln	Pro
Arg	Pro 530	Asp	His	Pro	Asp	Val 535	Pro	Ile	Phe	Pro	Gln 540	Pro	Glu	Arg	Ser
Met 545	Tyr	Arg	Gly	Gly	Glu 550	Phe	Gly	Tyr	Thr	Arg 555	Leu	Ala	Ala	Thr	Arg 560
Glu	Lys	Leu	Thr	Leu 565	Thr	Tyr	Val	Gly	Asn 570	His	Asp	Gly	Gln	Val 575	His
Asp	Ile	Met	Glu 580	Ile	Phe	Ser	Gly	Leu 585	Val	Ser	Ser	Asn	Ser 590	Ser	Val
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Arg Lys Ile Ser Pro Leu Tyr Leu Glu Ile Gly Gly Ser Val Leu Phe 610 620

Ala Leu Leu Gly Phe Ser Phe Gly Phe Leu Ile Arg Arg Lys 625 630 635 640

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<210> SEQ ID NO 30

<211> LENGTH: 1950

<212> TYPE: DNA

<213> ORGANISM: Solanum tuberosum

<400> SEQUENCE: 30

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Leu	Lys 610	Ile	Pro	Leu	Phe	Ser 615	Leu	Glu	Ile	Val	Gly 620	Ser	Val	Met	Phe
Ala 625	Leu	Val	Leu	Gly	Phe 630	Ser	Leu	Gly	Phe	Leu 635	Ile	Arg	Arg	Lys	Lys 640
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<sup>&</sup>lt;212> TYPE: DNA <213> ORGANISM: Gossypium hirsutum

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Pro Met Tyr Thr Thr Ser Phe Glu Ser Arg Asp Ala Pro 35 40 45	Leu Arg Glu
Lys Met Leu Glu His Leu Glu Pro Leu Phe Val Lys Asn 50 55 60	Asn Val Asn
Leu Ala Leu Trp Gly His Val His Arg Tyr Glu Arg Phe 65 70 75	Cys Pro Leu 80
Lys Asn Phe Thr Cys Gly Ser Met Gly Gln Lys Gly Lys 85 90	Asp Trp Glu 95
Ala Phe Pro Val His Val Val Ile Gly Met Ala Gly Gln	Asp Trp Gln
Pro Thr Trp Glu Pro Arg Pro Asp His Pro Thr Ile Pro	
Asn Pro Lys Arg Ser Leu Tyr Arg Thr Gly Glu Phe Gly	
Leu Ile Ala Thr Lys Glu Lys Leu Thr Leu Ser Phe Val	=
	160
Asp Gly Glu Val His Asp Met Val Glu Ile Leu Ala Ser 165 170	175
Leu Asn Gly Gly Asp Asp Asn Asn Gly Lys Val Gly Ala 180 185	Val His Lys 190
Val Asp Asp Val Thr Arg Tyr Ser Phe Ser His Tyr Val 195 200 205	
Ser Val Leu Val Leu Gly Gly Phe Val Gly Tyr Val Leu 210 215 220	Gly Phe Val

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Lys Thr Glu Glu Gln 245

<210> SEQ ID NO 38

<211> LENGTH: 930 <212> TYPE: DNA

<213> ORGANISM: Panicum virgatum

<400> SEQUENCE: 38

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<212> TYPE: PRT

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<400> SEQUENCE: 39

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Gly Asn Ser Val Leu Pro Thr Gly Asn Gly Gly Pro Asp Thr Arg Asn 35 \$40\$ 45

Leu Tyr Tyr Ser Phe Asp Ser Gly Val Val His Phe Val Tyr Met Ser 50 55 60

Thr Glu Thr Asn Phe Leu Gln Gly Ser Asp Gln Tyr Asn Phe Leu Lys 65 70 75 80

Ala Asp Leu Glu Lys Val Asn Arg Thr Arg Thr Pro Phe Val Val Phe

Gln Gly His Arg Pro Met Tyr Thr Ser Ser Asp Glu Thr Arg Asp Ala 100 105 110

Ala Leu Lys Gln Gln Met Leu Gln Asn Leu Glu Pro Leu Leu Val Thr \$115\$ \$120\$ \$125\$

Tyr Asn Val Thr Leu Ala Leu Trp Gly His Val His Arg Tyr Glu Arg 130 135 140

Phe Cys Pro Met Lys Asn Phe Gln Cys Val Asn Thr Ser Ser Ser Phe 145 150 155 160	
Gln Tyr Pro Gly Ala Pro Val His Leu Val Ile Gly Met Gly Gln 165 170 175	
Asp Trp Gln Pro Ile Trp Gln Pro Arg Pro Asp His Pro Asp Val Pro 180 185 190	
Ile Phe Pro Gln Pro Glu Arg Ser Met Tyr Arg Gly Gly Val Phe Gly 195 200 205	
Tyr Thr Arg Leu Val Ala Thr Arg Glu Lys Leu Thr Leu Thr Tyr Val 210 215 220	
Gly Asn His Asp Gly Gln Val His Asp Met Val Glu Ile Phe Ser Gly 225 230 235 240	
Gln Val Ser Ser Asn Ser Ser Val Ala Glu Ala Val Asp Gly Ala Lys 245 250 255	
Leu Ser Thr Gly Val Ser Thr Val Arg Lys Met Pro Pro Leu Tyr Leu 260 265 270	
Glu Ile Gly Gly Ser Val Met Phe Ala Leu Leu Leu Gly Phe Gly Phe 275 280 285	
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	120
cctgccctta tctcacatat tggagatatc agctacgcca gaggatactc ttggttgtgg	120 180
cctgccctta tctcacatat tggagatatc agctacgcca gaggatactc ttggttgtgg	
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115 116

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                                                                  240
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                              25
Leu Val Ala Thr Arg Glu Lys Leu Thr Leu Thr Tyr Val Gly Asn His
                         40
Asp Gly Gln Val His Gly Met Val Glu Ile Phe Ser Gly Leu Val Ser
Ser Asn Ser Ser Val Ala Val Ala Val His Asp Thr Lys Leu Gly Thr
Glu Val Ser Thr Val Arg Lys Ile Ser Pro Leu Tyr Leu Glu Ile Gly
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Glu Ser
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Ile Pro Tyr 35	Ser Val L	ys Phe Arg 40	Met Pro Gl	y Asn Ser Val 45	Leu Pro
Thr Gly Asn 50	Gly Ala P	ro Asp Thr 55	Arg Asn Le	ı Tyr Tyr Sei 60	Phe Asp
Ser Gly Val 65	Val His P	-	Met Ser Th	r Glu Thr Asr	n Phe Val 80
Gln Gly Ser	Asp Gln H 85	is Asn Phe	Leu Lys Al	a Asp Leu Glu	ı Lys Val 95
Asn Arg Ser	Arg Thr P	ro Phe Val	Val Phe Gl: 105	n Gly His Arg	,
Tyr Thr Ser 115		lu Ala Arg 120	Asp Phe Al	a Met Arg Glr 125	n Gln Met
Ile Gln His 130	Leu Glu L	eu Leu Leu 135	Val Met Ty	r Asn Val Thi 140	Leu Ala
Leu Trp Gly 145		is Arg Tyr 50	Glu Arg Ph	e Cys Pro Met 5	Lys Asn 160
Ser Gln Cys	Leu Asn T	nr Ser Ser	Ser Phe Il	e Tyr Pro Gly	Ala Pro 175
Val His Val	Val Ile G	ly Met Ala	Gly Gln As 185	p Trp Gln Pro	_
Gln Pro Arg		is Pro Asp 200		e Phe Pro Glr 205	n Pro Gly
Ile Ser Met 210	Tyr Arg G	ly Gly Glu 215	Phe Gly Ty	r Thr Lys Let 220	ı Val Ala
Thr Arg Glu 225	-	nr Leu Met 30	Tyr Val Gl	y Asn His Asp 5	Gly Gln 240
				n Thr Ser Thi	
	245		250	501 111	255

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Ser Ala Thr Glu Ala Val Asn Gln Thr Lys Leu Gly Ser Gly Thr Ser  $260 \hspace{1cm} 265 \hspace{1cm} 265 \hspace{1cm} 270 \hspace{1cm}$ 

Met Leu Ala Leu Leu Leu Gly Phe Ala Leu Gly Phe Leu Leu Arg Lys 290 295 300

Lys Arg Glu Ala Ala Gln Trp Thr Pro Val Lys Asn Glu Glu Ser 305  $\phantom{\bigg|}$  310  $\phantom{\bigg|}$  315

<210> SEQ ID NO 46

<211> LENGTH: 1959

<212> TYPE: DNA

<213> ORGANISM: Brassica napus

<400> SEQUENCE: 46

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caaagcaa	ccttatccat	ctcccccaaa	actctaagcc	gatccggcga	ttccatcctc	120
caaatggt	ccaacgtcga	ctctccctcc	gatctcgact	ggctaggcat	ctactccccc	180
agactctc	cccacgacca	cttcatcggc	tacaaattcc	tcaacgtctc	ccccacgtgg	240
atccggct	ccggcgcgat	ctccctcccc	ctcaccaacc	tccgatcgaa	ctacacgttc	300
tatcttcc	gatggacgca	gtccgagatc	aatccgaagc	acaaggacca	cgaccagaat	360
cttaccgg	gaacgaagca	ccttctggcg	gaatcggagc	aggtggggtt	cggatccgcc	420
cgtgggga	ggccggagca	gatccatttg	gcgttcgagg	ataaggttaa	caggatgcgg	480
cacgttcg	tagctgggga	tggggaagaa	aggttcgtga	ggtacggaga	ggggaaggac	540
gttggcga	actccgcggc	ggegegeggg	attaggtacg	agagggagca	tatgtgtaat	600
teeggeta	attccaccgt	gggatggaga	gatcccgggt	ggatttttca	taccgttatg	660
gaatttga	acggtggcgt	taggtattat	tatcaggttg	ggagtgattc	aaagggatgg	720
tgagatcc	acagctttat	cgctcgagat	atctactcag	aagaaaccat	agctttcatg	780
cggagaca	tgggttgcgc	tacaccttac	aataccttta	teeggaegea	ggacgagagt	840
ctcaacag	ttaagtggat	actccgcgac	atcgaagctc	ttggtgacaa	gccagctctt	900
ttcgcaca	ttggtgatat	aagctacgct	cgtggttact	cgtgggtgtg	ggatgagttc	960
cgctcaga	tcgagcctat	tgcctcgaga	gttccttacc	acgtctgcat	tggtaaccac	1020
gtatgact	tccctactca	gccgtggaaa	cctgattggg	gaacttacgg	taatgacggt	1080
gggagagt	gcggtgtgcc	gtatagtete	aagttcaaca	tgcctggaaa	ctcgtcggaa	1140
aacgggaa	cgaaagctcc	tcctacaagg	aatttgtatt	actcttacga	catggggtcg	1200
tcatttcc	tttacatctc	caccgagacg	aactttctca	aaggagggag	gcaatacgag	1260
tataaagc	gagatettga	gtctgtgaac	agggagaaaa	caccgtttgt	tgtcgtgcaa	1320
acacagac	cgatgtacac	cacgagcaac	gaggtgagag	acgcgatgat	taggcaaaag	1380
ggtggagc	atttggagcc	gctgtttgtg	gagaacaacg	tgacgcttgc	tctgtgggga	1440
tgttcata	gatacgagag	gttttgtccg	ataagcaaca	acacgtgtgg	gaaacagtgg	1500
aggaagcc	cggttcatct	tgtgatcggt	atgggcggtc	aagactggca	accgatttgg	1560
gccgagac	cgaaccatcc	gggtcttcct	atattccctc	agcctgaaca	gtcgatgtac	1620
gacgggtg	agtttgggta	cactcgtttg	gttgcgaaca	aagagaagct	tactgtttcg	1680
tgtgggta	accatgatgg	agaagttcat	gatagtgttg	agatettage	gtctggggaa	1740

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gtaatcagtg ggaggaaaga ggaaactatt aagaccgttc ctgtatctgc aacacttgtg gggaaacctg agtctgatgt cttatggtat gttaaaggag caggcttgtt ggttatgggt gtgcttttag ggttccttat agggtttttt acaaggggga agaaaggatc ttcttcatct gataaccgtt ggatcccagt caagaacgag gagacatga 1959 <210> SEQ ID NO 47 <211> LENGTH: 652 <212> TYPE: PRT <213> ORGANISM: Brassica napus <400> SEQUENCE: 47 Met Ile Val Asp Phe Ser Thr Phe Ile Leu Phe Ile Ser Val Phe Ile Ser Ser Ala Asn Ala Lys Ala Thr Leu Ser Ile Ser Pro Lys Thr Leu Ser Arg Ser Gly Asp Ser Ile Leu Ile Lys Trp Ser Asn Val Asp Ser Pro Ser Asp Leu Asp Trp Leu Gly Ile Tyr Ser Pro Pro Asp Ser Pro His Asp His Phe Ile Gly Tyr Lys Phe Leu Asn Val Ser Pro Thr Trp 65 70 75 80 Gln Ser Gly Ser Gly Ala Ile Ser Leu Pro Leu Thr Asn Leu Arg Ser Asn Tyr Thr Phe Arg Ile Phe Arg Trp Thr Gln Ser Glu Ile Asn Pro 105 Lys His Lys Asp His Asp Gln Asn Pro Leu Pro Gly Thr Lys His Leu 120 Leu Ala Glu Ser Glu Gln Val Gly Phe Gly Ser Ala Gly Val Gly Arg Pro Glu Gln Ile His Leu Ala Phe Glu Asp Lys Val Asn Arg Met Arg Val Thr Phe Val Ala Gly Asp Gly Glu Glu Arg Phe Val Arg Tyr Gly Glu Gly Lys Asp Ala Leu Ala Asn Ser Ala Ala Ala Arg Gly Ile Arg Tyr Glu Arg Glu His Met Cys Asn Ala Pro Ala Asn Ser Thr Val Gly Trp Arg Asp Pro Gly Trp Ile Phe His Thr Val Met Lys Asn Leu Asn Gly Gly Val Arg Tyr Tyr Gln Val Gly Ser Asp Ser Lys Gly Trp Ser Glu Ile His Ser Phe Ile Ala Arg Asp Ile Tyr Ser Glu Glu Thr  $245 \hspace{1cm} 250 \hspace{1cm} 255$ Ile Ala Phe Met Phe Gly Asp Met Gly Cys Ala Thr Pro Tyr Asn Thr 265 Phe Ile Arg Thr Gln Asp Glu Ser Ile Ser Thr Val Lys Trp Ile Leu Arg Asp Ile Glu Ala Leu Gly Asp Lys Pro Ala Leu Val Ser His Ile Gly Asp Ile Ser Tyr Ala Arg Gly Tyr Ser Trp Val Trp Asp Glu Phe 310 315 Phe Ala Gln Ile Glu Pro Ile Ala Ser Arg Val Pro Tyr His Val Cys

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Ile Gly Asn His Glu Tyr Asp Phe Pro Thr Gln Pro Trp Lys Pro Asp
Trp Gly Thr Tyr Gly Asn Asp Gly Gly Glu Cys Gly Val Pro Tyr
Ser Leu Lys Phe Asn Met Pro Gly Asn Ser Ser Glu Pro Thr Gly Thr
Lys Ala Pro Pro Thr Arg Asn Leu Tyr Tyr Ser Tyr Asp Met Gly Ser
Val His Phe Leu Tyr Ile Ser Thr Glu Thr Asn Phe Leu Lys Gly Gly
 \hbox{Arg Gln Tyr Glu Phe Ile Lys Arg Asp Leu Glu Ser Val Asn Arg Glu } \\
Lys Thr Pro Phe Val Val Val Gln Gly His Arg Pro Met Tyr Thr Thr
Ser Asn Glu Val Arg Asp Ala Met Ile Arg Gln Lys Met Val Glu His
Leu Glu Pro Leu Phe Val Glu Asn Asn Val Thr Leu Ala Leu Trp Gly 465 470 475 480
His Val His Arg Tyr Glu Arg Phe Cys Pro Ile Ser Asn Asn Thr Cys
                                  490
Gly Lys Gln Trp Arg Gly Ser Pro Val His Leu Val Ile Gly Met Gly
                      505
Gly Gln Asp Trp Gln Pro Ile Trp Gln Pro Arg Pro Asn His Pro Gly
                          520
Leu Pro Ile Phe Pro Gln Pro Glu Gln Ser Met Tyr Arg Thr Gly Glu
                       535
Phe Gly Tyr Thr Arg Leu Val Ala Asn Lys Glu Lys Leu Thr Val Ser
                   550
                                        555
Phe Val Gly Asn His Asp Gly Glu Val His Asp Ser Val Glu Ile Leu
                                  570
Ala Ser Gly Glu Val Ile Ser Gly Arg Lys Glu Glu Thr Ile Lys Thr
                                585
Val Pro Val Ser Ala Thr Leu Val Gly Lys Pro Glu Ser Asp Val Leu
Trp Tyr Val Lys Gly Ala Gly Leu Leu Val Met Gly Val Leu Leu Gly
Phe Leu Ile Gly Phe Phe Thr Arg Gly Lys Lys Gly Ser Ser Ser
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<223 > OTHER INFORMATION: Xaa is any amino acid
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<212> TYPE: PRT
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Xaa Asp Xaa Xaa Tyr
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Gly Asn His Glu
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<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
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<223> OTHER INFORMATION: Xaa is Leu, Ile, Val, Phe, or Met
<220> FEATURE:
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<222> LOCATION: (2)..(2)
<223 > OTHER INFORMATION: Xaa is any amino acid
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<222> LOCATION: (3)..(3)
<223> OTHER INFORMATION: Xaa is any amino acid
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Xaa His Xaa His
1
<210> SEQ ID NO 54
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<212> TYPE: PRT
<213> ORGANISM: Arabidopsis thaliana
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<222> LOCATION: (12)..(12)
<223> OTHER INFORMATION: Xaa is Phe or Trp
<400> SEQUENCE: 54
Tyr His Val Cys Ile Gly Asn His Glu Tyr Asp Xaa
<210> SEQ ID NO 55
<211> LENGTH: 12
<212> TYPE: PRT
<213 > ORGANISM: Artificial Sequence
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<222> LOCATION: (12)..(12)
<223> OTHER INFORMATION: Xaa is Phe or Trp
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<210> SEQ ID NO 56
<211> LENGTH: 13
<212> TYPE: PRT
<213> ORGANISM: Arabidopsis thaliana
<400> SEQUENCE: 56
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<210> SEQ ID NO 57
<211> LENGTH: 18
<212> TYPE: PRT
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<400> SEQUENCE: 57
Lys Glu Lys Leu Thr Val Ser Phe Val Gly Asn His Asp Gly Glu Val
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<210> SEQ ID NO 58
<211> LENGTH: 18
<212> TYPE: PRT
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<220> FEATURE:
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                                  10
His Asp
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Lys Glu Lys Leu Thr Leu Thr Tyr Ile Gly Asn His Asp Gly Gln Val
His Asp
<210> SEQ ID NO 61
<211> LENGTH: 10
<212> TYPE: PRT
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<222> LOCATION: (8)..(9)
<223> OTHER INFORMATION: Xaa is any amino acid
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                5
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1 5
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<211> LENGTH: 23
<212> TYPE: PRT
<213 > ORGANISM: Arabidopsis thaliana
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Gly Phe Ile Ile Gly Phe Phe
            20
<210> SEO ID NO 66
<211> LENGTH: 12
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
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<223> OTHER INFORMATION: Xaa is Leu, M, or V
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
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<223> OTHER INFORMATION: Xaa is Leu, Ile, Val, Phe, or Met
<220> FEATURE:
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<223> OTHER INFORMATION: Xaa is Gly or Ala
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<221> NAME/KEY: MISC_FEATURE
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<223> OTHER INFORMATION: Xaa is Val, Ala, or Leu
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<221> NAME/KEY: MISC_FEATURE
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<223> OTHER INFORMATION: Xaa is Leu, Ile, Val, Phe, or Met
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<221> NAME/KEY: MISC_FEATURE
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<223> OTHER INFORMATION: Xaa is Phe or Tyr
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<223> OTHER INFORMATION: Xaa is any amino acid residue
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<223> OTHER INFORMATION: Xaa is Leu, Ile, Val, Phe, or Met
<400> SEQUENCE: 66
Xaa Xaa Xaa Xaa Xaa Xaa Gly Xaa Xaa Gly
<210> SEQ ID NO 67
<211> LENGTH: 529
<212> TYPE: PRT
```

-212	0 - OI	ייי אייי	CM.	7 mal	ad dor	aia	+b a l	ione							
					olaog	psis	thal	llana	ı						
< 400	O> SI	EQUE	ICE :	67											
Met 1	Thr	Phe	Leu	Leu 5	Leu	Leu	Leu	Phe	Cys	Phe	Leu	Ser	Pro	Ala 15	Ile
Ser	Ser	Ala	His 20	Ser	Ile	Pro	Ser	Thr 25	Leu	Asp	Gly	Pro	Phe 30	Val	Pro
Val	Thr	Val 35	Pro	Leu	Asp	Thr	Ser 40	Leu	Arg	Gly	Gln	Ala 45	Ile	Asp	Leu
Pro	Asp 50	Thr	Asp	Pro	Arg	Val 55	Arg	Arg	Arg	Val	Ile 60	Gly	Phe	Glu	Pro
Glu 65	Gln	Ile	Ser	Leu	Ser 70	Leu	Ser	Ser	Asp	His 75	Asp	Ser	Ile	Trp	Val 80
Ser	Trp	Ile	Thr	Gly 85	Glu	Phe	Gln	Ile	Gly 90	Lys	Lys	Val	Lys	Pro 95	Leu
Asp	Pro	Thr	Ser 100	Ile	Asn	Ser	Val	Val 105	Gln	Phe	Gly	Thr	Leu 110	Arg	His
Ser	Leu	Ser 115	His	Glu	Ala	ГЛа	Gly 120	His	Ser	Leu	Val	Tyr 125	Ser	Gln	Leu
Tyr	Pro 130	Phe	Asp	Gly	Leu	Leu 135	Asn	Tyr	Thr	Ser	Gly 140	Ile	Ile	His	His
Val 145	Arg	Ile	Thr	Gly	Leu 150	Lys	Pro	Ser	Thr	Ile 155	Tyr	Tyr	Tyr	Arg	Cys 160
Gly	Asp	Pro	Ser	Arg 165	Arg	Ala	Met	Ser	Lys 170	Ile	His	His	Phe	Arg 175	Thr
Met	Pro	Val	Ser 180	Ser	Pro	Ser	Ser	Tyr 185	Pro	Gly	Arg	Ile	Ala 190	Val	Val
Gly	Asp	Leu 195	Gly	Leu	Thr	Tyr	Asn 200	Thr	Thr	Asp	Thr	Ile 205	Ser	His	Leu
Ile	His 210	Asn	Ser	Pro	Asp	Leu 215	Ile	Leu	Leu	Ile	Gly 220	Asp	Val	Ser	Tyr
Ala 225	Asn	Leu	Tyr	Leu	Thr 230	Asn	Gly	Thr	Ser	Ser 235	Asp	Cya	Tyr	Ser	Cys 240
Ser	Phe	Pro	Glu	Thr 245	Pro	Ile	His	Glu	Thr 250	Tyr	Gln	Pro	Arg	Trp 255	Asp
Tyr	Trp	Gly	Arg 260	Phe	Met	Glu	Asn	Leu 265	Thr	Ser	Lys	Val	Pro 270	Leu	Met
Val	Ile	Glu 275	Gly	Asn	His	Glu	Ile 280	Glu	Leu	Gln	Ala	Glu 285	Asn	Lys	Thr
Phe	Glu 290	Ala	Tyr	Ser	Ser	Arg 295	Phe	Ala	Phe	Pro	Phe 300	Asn	Glu	Ser	Gly
Ser 305	Ser	Ser	Thr	Leu	Tyr 310	Tyr	Ser	Phe	Asn	Ala 315	Gly	Gly	Ile	His	Phe 320
Val	Met	Leu	Gly	Ala 325	Tyr	Ile	Ala	Tyr	330	Lys	Ser	Ala	Glu	Gln 335	Tyr
Glu	Trp	Leu	Lys 340	Lys	Asp	Leu	Ala	Lys 345	Val	Asp	Arg	Ser	Val 350	Thr	Pro
Trp	Leu	Val 355	Ala	Ser	Trp	His	Pro 360	Pro	Trp	Tyr	Ser	Ser 365	Tyr	Thr	Ala
His	Tyr 370	Arg	Glu	Ala	Glu	Сув 375	Met	Lys	Glu	Ala	Met 380	Glu	Glu	Leu	Leu
Tyr 385	Ser	Tyr	Gly	Thr	Asp 390	Ile	Val	Phe	Asn	Gly 395	His	Val	His	Ala	Tyr 400

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Glu	Arg	Ser	Asn	Arg 405	Val	Tyr	Asn	Tyr	Glu 410	Leu	Asp	Pro	Сув	Gly 415	Pro						
Val	Tyr	Ile	Val 420	Ile	Gly	Asp	Gly	Gly 425	Asn	Arg	Glu	Lys	Met 430	Ala	Ile						
Glu	His	Ala 435	Asp	Asp	Pro	Gly	Lys 440	Сла	Pro	Glu	Pro	Leu 445	Thr	Thr	Pro						
Asp	Pro 450	Val	Met	Gly	Gly	Phe 455	CÀa	Ala	Trp	Asn	Phe 460	Thr	Pro	Ser	Asp						
Lys 465	Phe	Сув	Trp	Asp	Arg 470	Gln	Pro	Asp	Tyr	Ser 475	Ala	Leu	Arg	Glu	Ser 480						
Ser	Phe	Gly	His	Gly 485	Ile	Leu	Glu	Met	Lys 490	Asn	Glu	Thr	Trp	Ala 495	Leu						
Trp	Thr	Trp	Tyr 500	Arg	Asn	Gln	Asp	Ser 505	Ser	Ser	Glu	Val	Gly 510	Asp	Gln						
Ile	Tyr	Ile 515	Val	Arg	Gln	Pro	Asp 520	Arg	Cys	Pro	Leu	His 525	His	Arg	Leu						
Val																					

What is claimed is:

- 1. A method to make a transgenic plant comprising: introducing a phosphatase gene into a plant, the phosphatase gene encoding a polypeptide comprising SEQ ID NO: 2 or a polypeptide having at least 90% sequence identity to SEQ ID NO: 2; SEQ ID NO: 2, wherein introducing the phosphatase gene into the plant increases growth rate, sugar content, crop yield, or the combination thereof, relative to wild-type plants.
- 2. The method of claim 1, wherein the phosphatase gene 35 is a purple acid phosphatase gene comprising a nucleotide sequence encoding a polypeptide comprising the amino acid sequence of SEQ ID NO: 2.
- 3. The method of claim 1, wherein the phosphatase gene is a purple acid phosphatase gene comprising the nucleotide 40 sequence of SEQ ID NO:1 or a nucleotide sequence having at least 90% sequence identity to SEQ ID NO:1.
- **4**. The method of claim **2**, wherein said nucleotide sequence comprises SEQ ID NO:1.
- 5. The method of claim 1, wherein introducing the phosphatase gene up-regulates the enzymatic activity of sucrose phosphate synthase in the transgenic plant relative to the wild-type plant.

- 6. The method of claim 1, wherein introducing the phosphatase gene up-regulates the sucrose and/or glucose level in the transgenic plant relative to the wild-type plant.
- 7. The method of claim 1, wherein introducing the phosphatase gene increases the growth rate of the transgenic plant relative to the wild-type plants.
- 8. The method of claim 1, wherein introducing the phosphatase gene results in a higher crop yield of the transgenic plant relative to the wild-type plants.
- 9. The method of claim 1, wherein the plant is a species selected from one of the group consisting of the following genera: Asparagus, Bromus, Hemerocalli, Hordeum, Loliu, Panicum, Pennisetum, Saccharum, Sorghum, Trigonell, Triticum, Zea, Antirrhinum, Arabidopsis, Arachis, Atropa, Brassica, Browallia, Capsicum, Carthamus, Cichorium, Citrus, Chrysanthemum, Cucumis, Datura, Daucus, Digitalis, Fragaria, Geranium, Glycine, Helianthus, Hyscyamus, Ipomoea, Latuca, Linum, Lotus, Solanum lycopersicon, Majorana, Malva, Gossypium, Manihot, Medicago, Nemesia, Nicotiana, Onobrychis, Pelargonium, Petunia, Ranunculus, Raphanus, Salpiglossis, Senecio, Sinapis, Solanum, Trifolium, Vigna, and Vitis.
- 10. The method of claim 1, wherein the plant is a species selected from the family *Brassica*.

\* \* \* \* \*